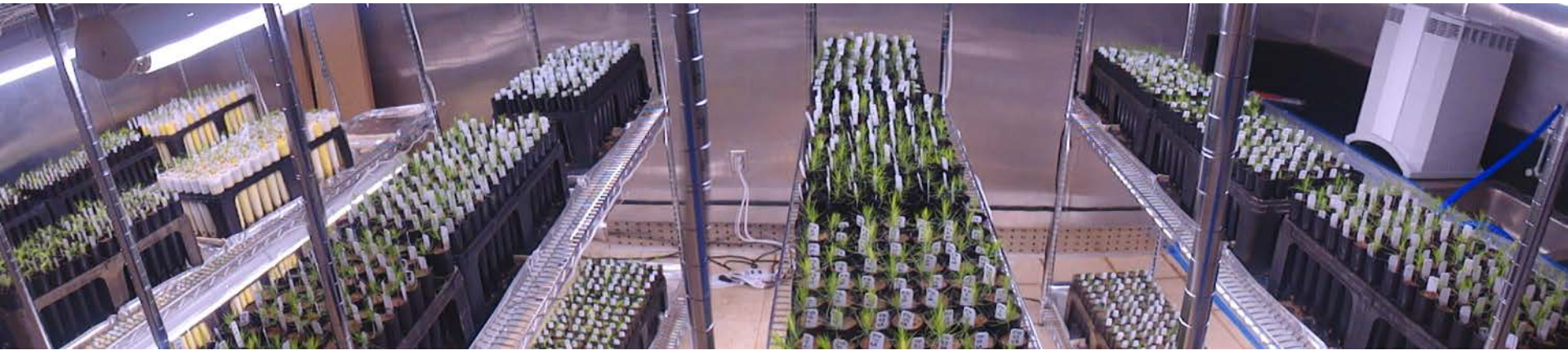


Fungal pathogen tolerance and geographic variation influence ectomycorrhizal traits of loblolly pine

B.J. Piculell, L.G. Eckhardt, J.D. Hoeksema

1. Department of Biology, P.O. Box 1848, University of Mississippi, University, MS 38677
2. Department of Biology, 66 George Street, College of Charleston, Charleston, SC 29424
3. School of Forestry and Wildlife Sciences, 602 Duncan Drive Suite 3301, Auburn University, Auburn, AL 36849



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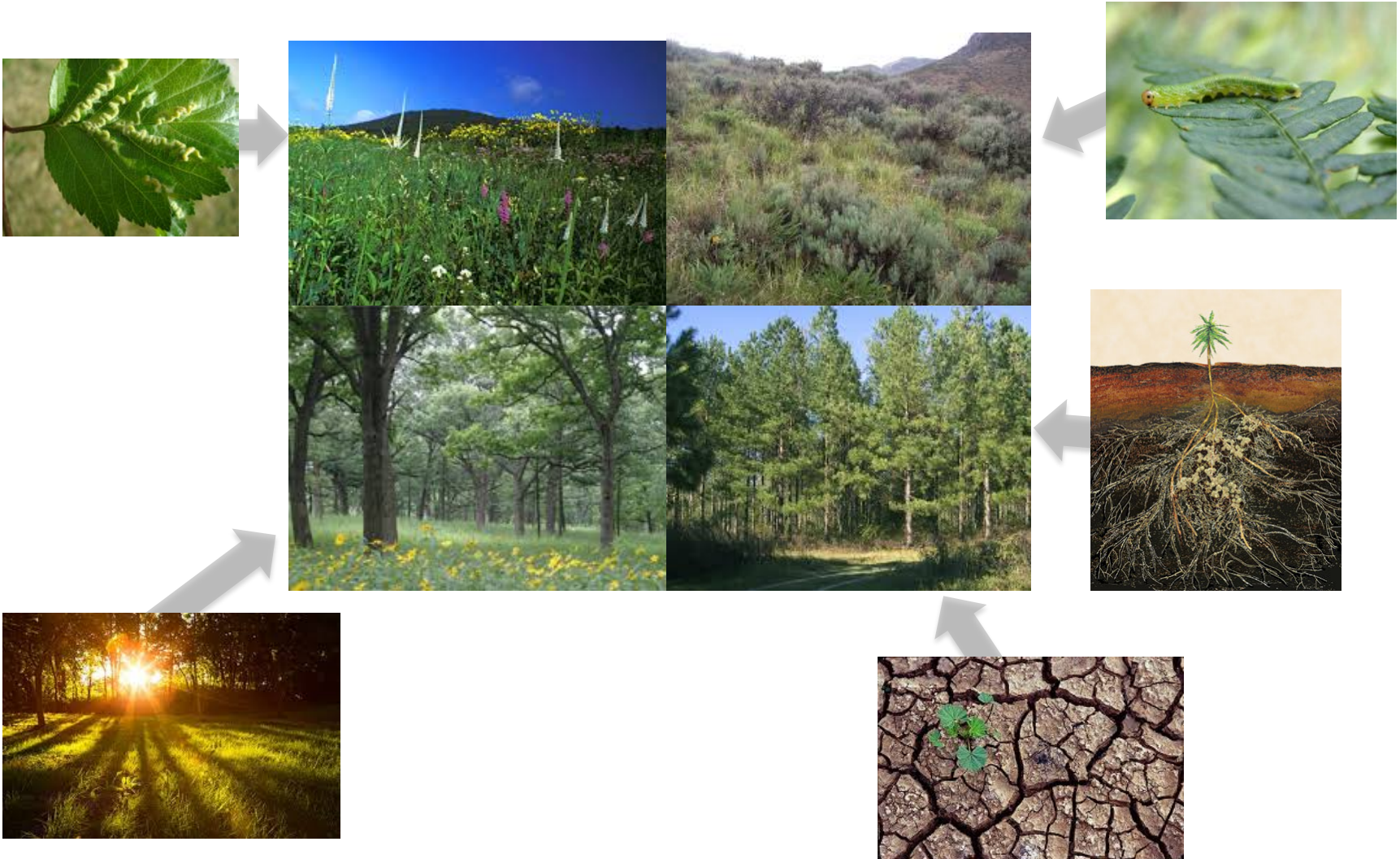


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How do plants evolve in response to complex environments?



How do plants evolve in response to complex environments?

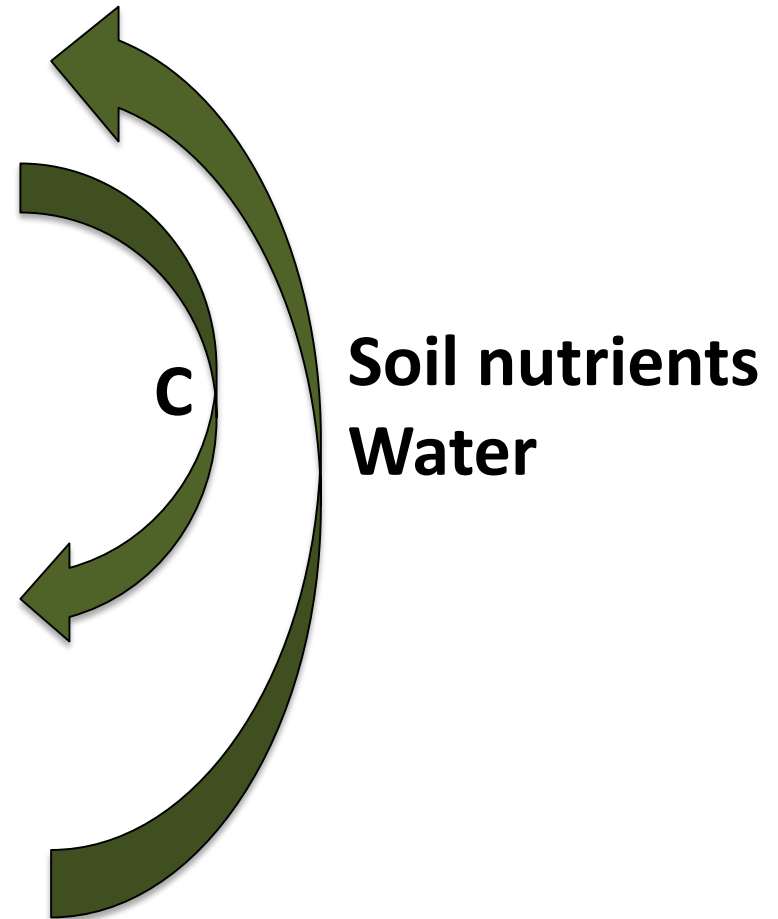
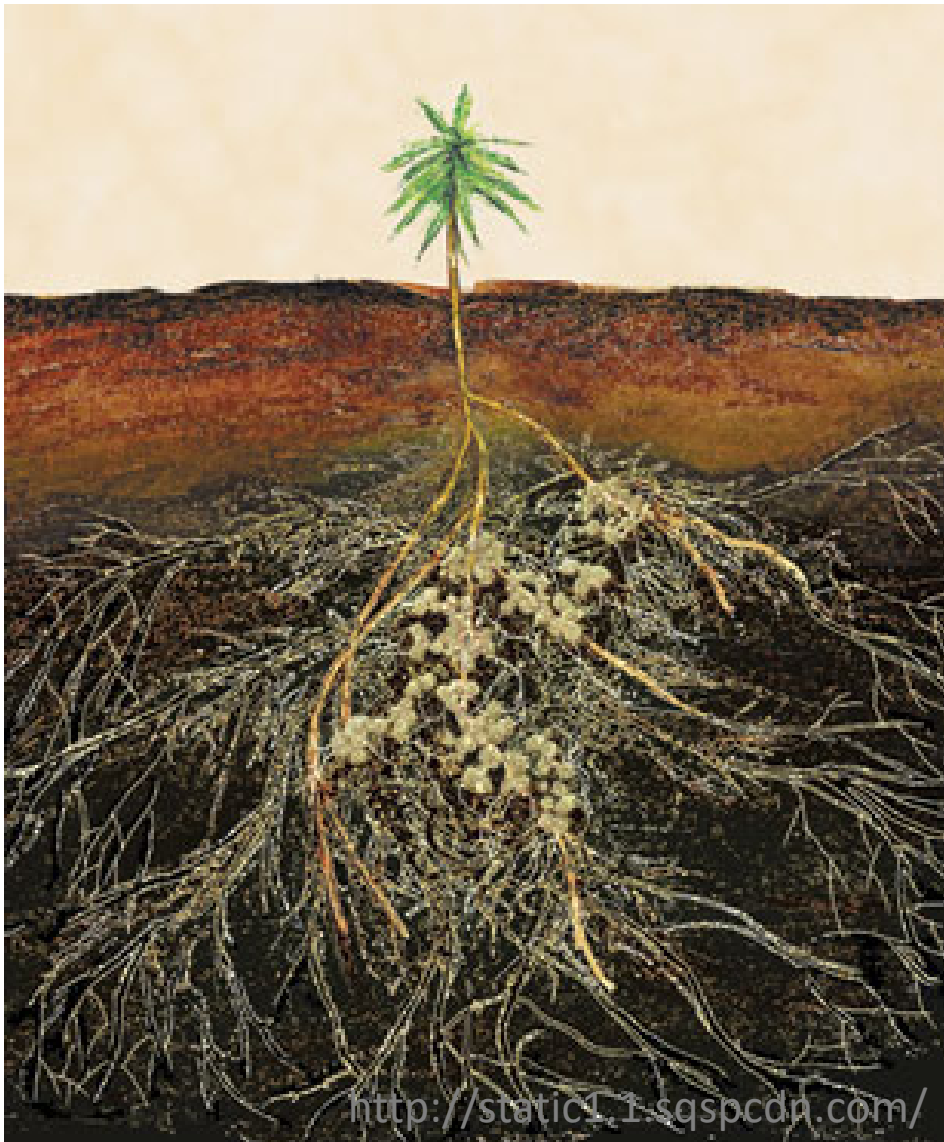




Loblolly pine (*Pinus taeda*)



mycorrhizal fungi

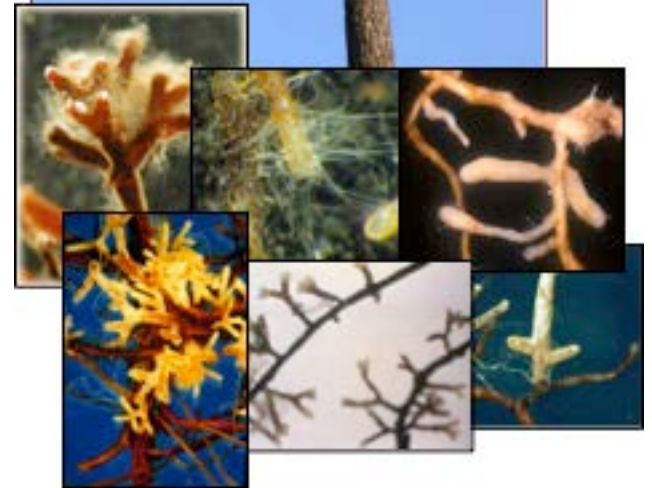


Ectomycorrhizal fungi ⇔ Plants

Alter competitive interactions
among plants

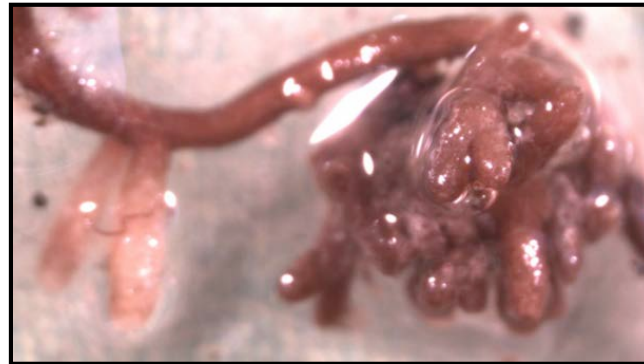
Multiple fungal species per host
plant

Vary in benefit to host plant



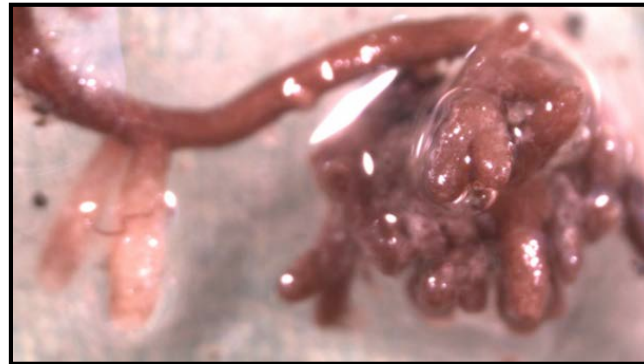
Morphotyping and fungal identification

Root tips colonized by ectomycorrhizal fungi grouped by similar phenotypic characteristics (color, texture, emanating hyphae)



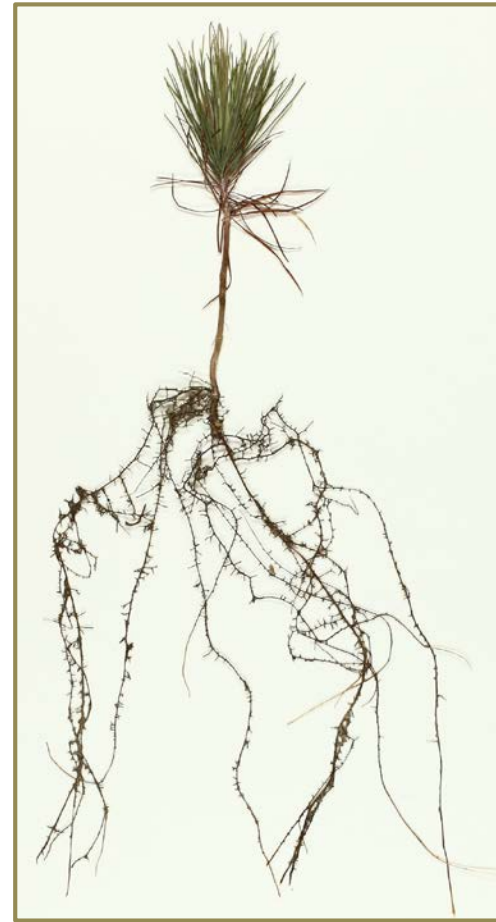
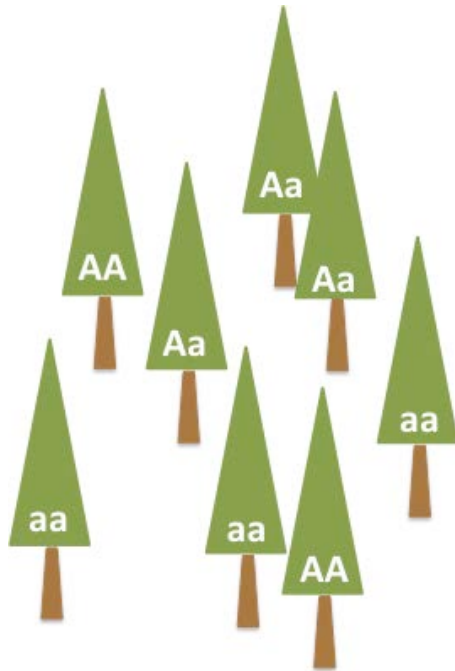
Morphotyping and fungal identification

Root tips colonized by ectomycorrhizal fungi grouped by similar phenotypic characteristics (color, texture, emanating hyphae)





Understanding the genetic basis of mycorrhizal traits





Evidence for genetic correlations between mycorrhizal and other plant traits



Modern wheat vs older cultivars
(Zhu *et al.*, 2001)



Cultivated tomato vs wild strains
(Bryla & Koide, 1990)



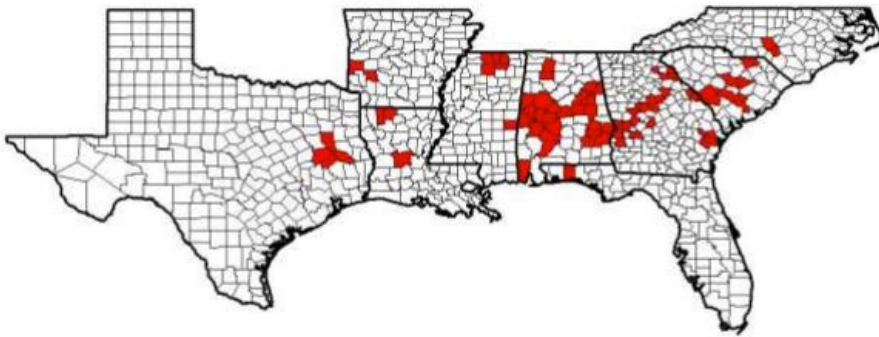
Herbivore resistant Pinyon pine
(Sthultz *et al.*, 2009)

Pine decline

Leptographium and *Grosmannia*



from Eckhardt *et al.*, 2007



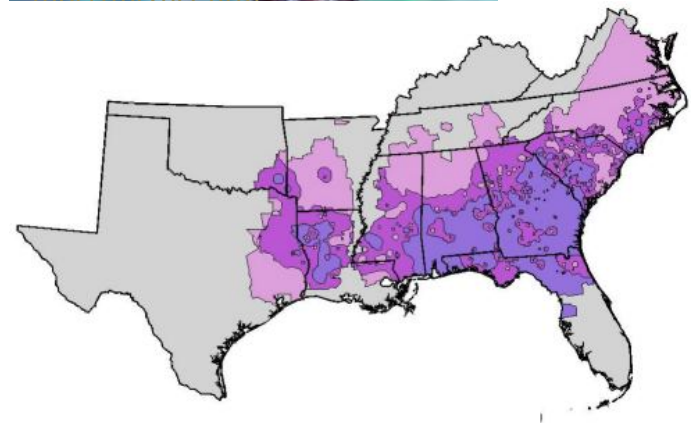
Fusiform rust

Cronartium



[/hosf/fusrust.htm](http://hosf/fusrust.htm)

[/hosf/fusrust.htm](http://hosf/fusrust.htm)





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Review

A review of southern pine decline in North America

David R. Coyle^{a,*}, Kier D. Klepzig^b, Frank H. Koch^c, Lawrence A. Morris^a, John T. Nowak^d, Steven W. Oak^{d,1}, William J. Orosina^{e,1}, William D. Smith^{c,1}, Kamal J.K. Gandhi^a







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Managing Fusiform Rust on Loblolly and Slash Pine in Forest and Landscape Settings

GEORGIA FORESTRY COMMISSION




gatrees.org

Loblolly Pine Decline

James Johnson, Forest Health Coordinator

A phenomenon known as Loblolly Pine (*Pinus taeda*) Decline is now used to describe drastic decreases in tree health, growth rates, and ongoing mortality in stands. Stands suffering from LPD are characterized by thinning and yellowing crowns, reduced radial growth, deterioration of fine feeder root systems, and increased mortality rates on



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University of Arkansas System

Agriculture and Natural Resources

FSA7543

Fusiform Rust in Arkansas

ASSESSMENT OF LOBLOLLY PINE DECLINE IN CENTRAL ALABAMA

Nolan J. Hess, William J. Orosina, Emily A. Carter, Jim R. Steinman, John P. Jones, Lori G. Eckhardt, Ann M. Weber, and Charles H. Walkinshaw¹

Abstract—Loblolly pine (*Pinus taeda* L.) decline has been prevalent on upland sites of central Alabama since the 1960's. The purpose of this study was to compare Forest Health Monitoring (FHM) standards and protocols with root health evaluations relative to crown, stem, and site measurements. Thirty-nine 1/6 acre plots were established on loblolly decline sites in nine central Alabama counties. Sites were selected on federal, state, and private industrial lands to measure variables of decline symptoms, age classes and management

Fungal pathogen tolerance and geographic variation influence ectomycorrhizal traits of loblolly pine

Q1: Do different locations within the natural range of loblolly pine yield different mycorrhizal fungal communities?

Q2: Do individual EM fungi respond differently to host genetic variation in pathogen tolerance and does this response depend on origin of the fungal community?



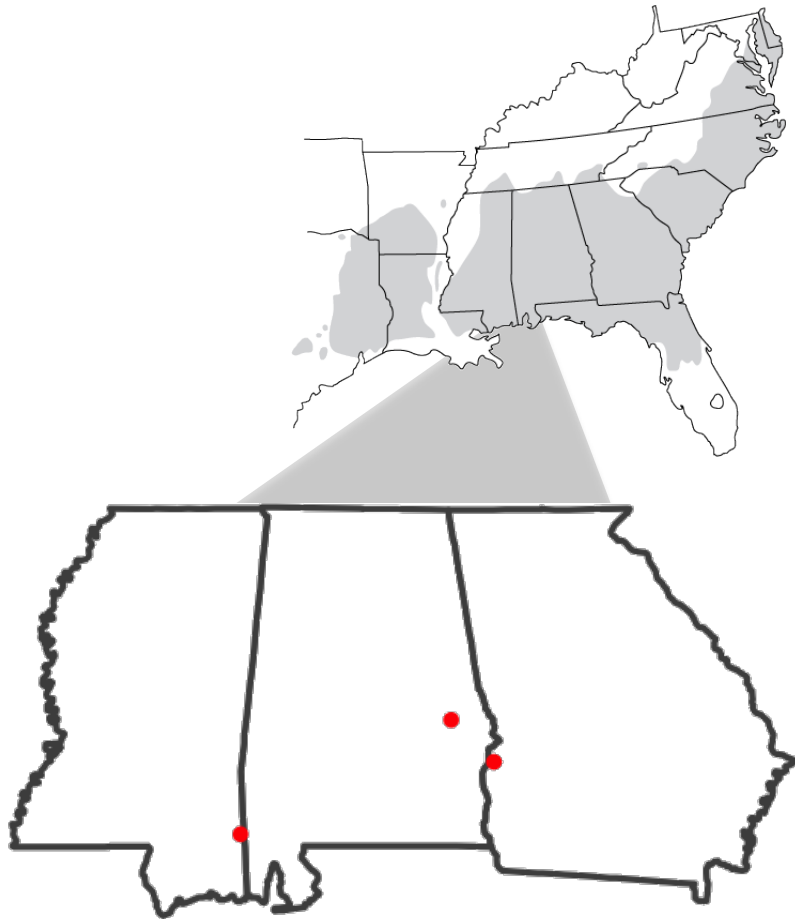
Seedlings from loblolly pine families either *tolerant* or *susceptible* to Pine decline (PD) or Fusiform rust (FR)

Tolerant

Susceptible



3 soil sources





seedlings: 4 categories

Fusiform rust: t or s

Pine decline: t or s

Category	# Individual seedlings		
	AL	MS	GA
FR-t	52	42	37
FR-s	63	54	55
PD-t	48	50	23
PD-s	50	43	41
Total	213	189	156



Fungal pathogen tolerance and geographic variation influence ectomycorrhizal traits of loblolly pine

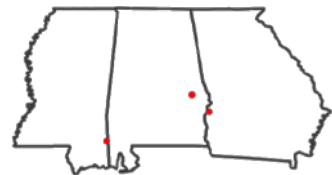
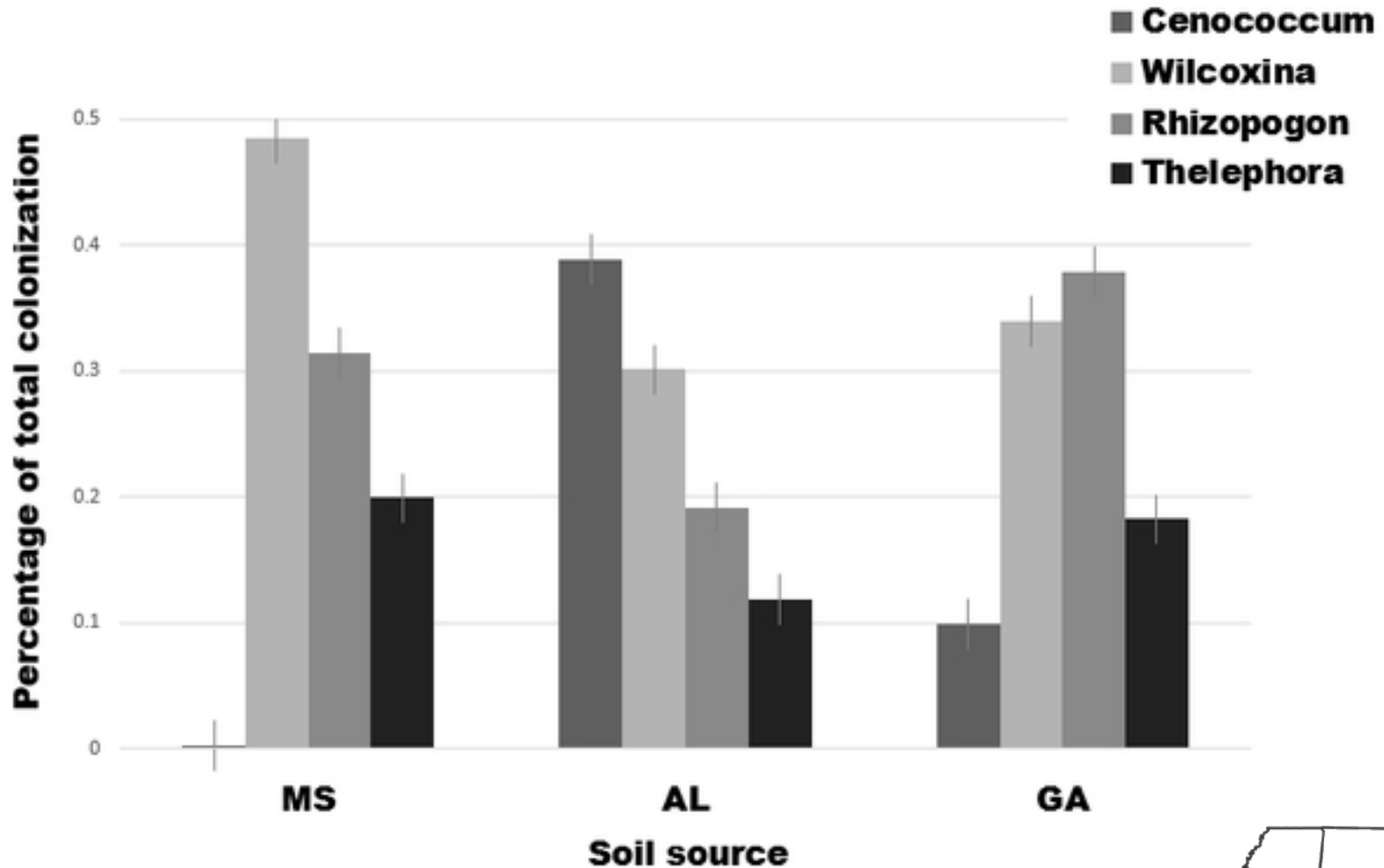
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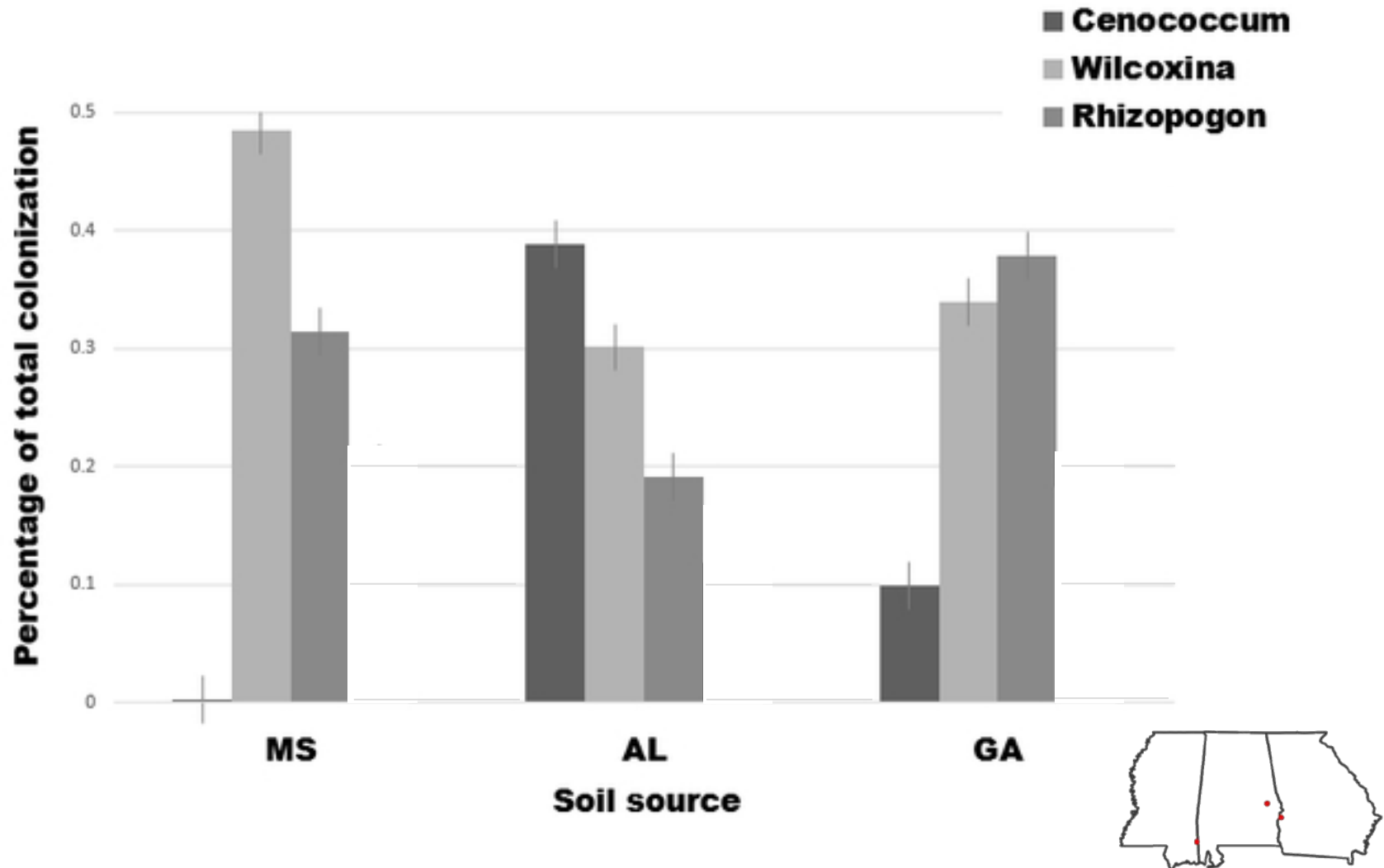
Different locations within the natural range of loblolly pine yield different dominant mycorrhizal fungal communities

($F_{2,560} = 36.754$, $p = 0.01$)



Three of the four dominant fungal colonizers respond only to soil inoculation source

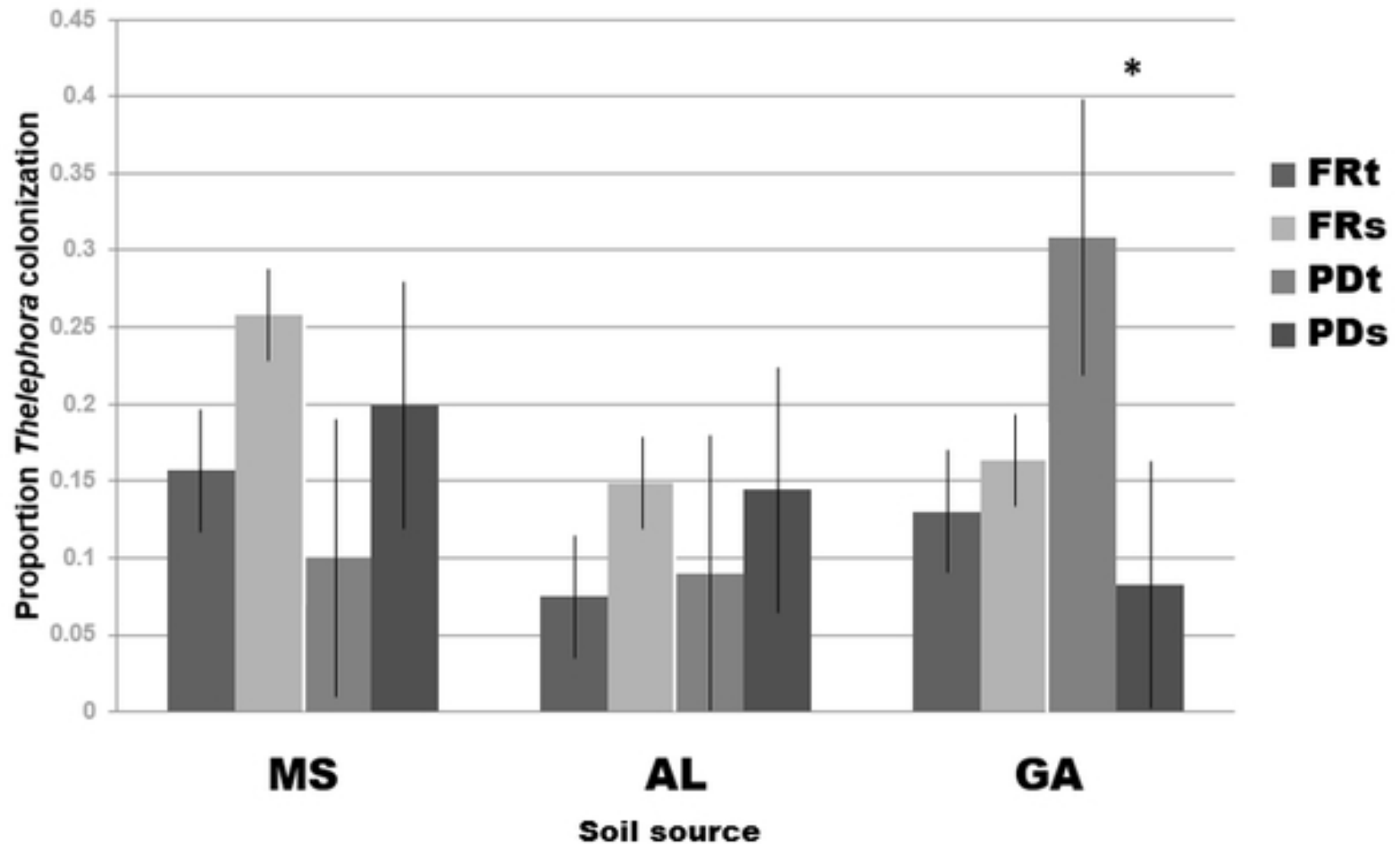
Rhizopogon ($F_{2,532} = 19.342$, $p = 7.787 \times 10^{-9}$), *Cenococcum* ($F_{2,532} = 120.840$, $p < 2.0 \times 10^{-16}$), and *Thelephora* ($F_{2,532} = 12.5084$, $p = 4.91 \times 10^{-6}$)





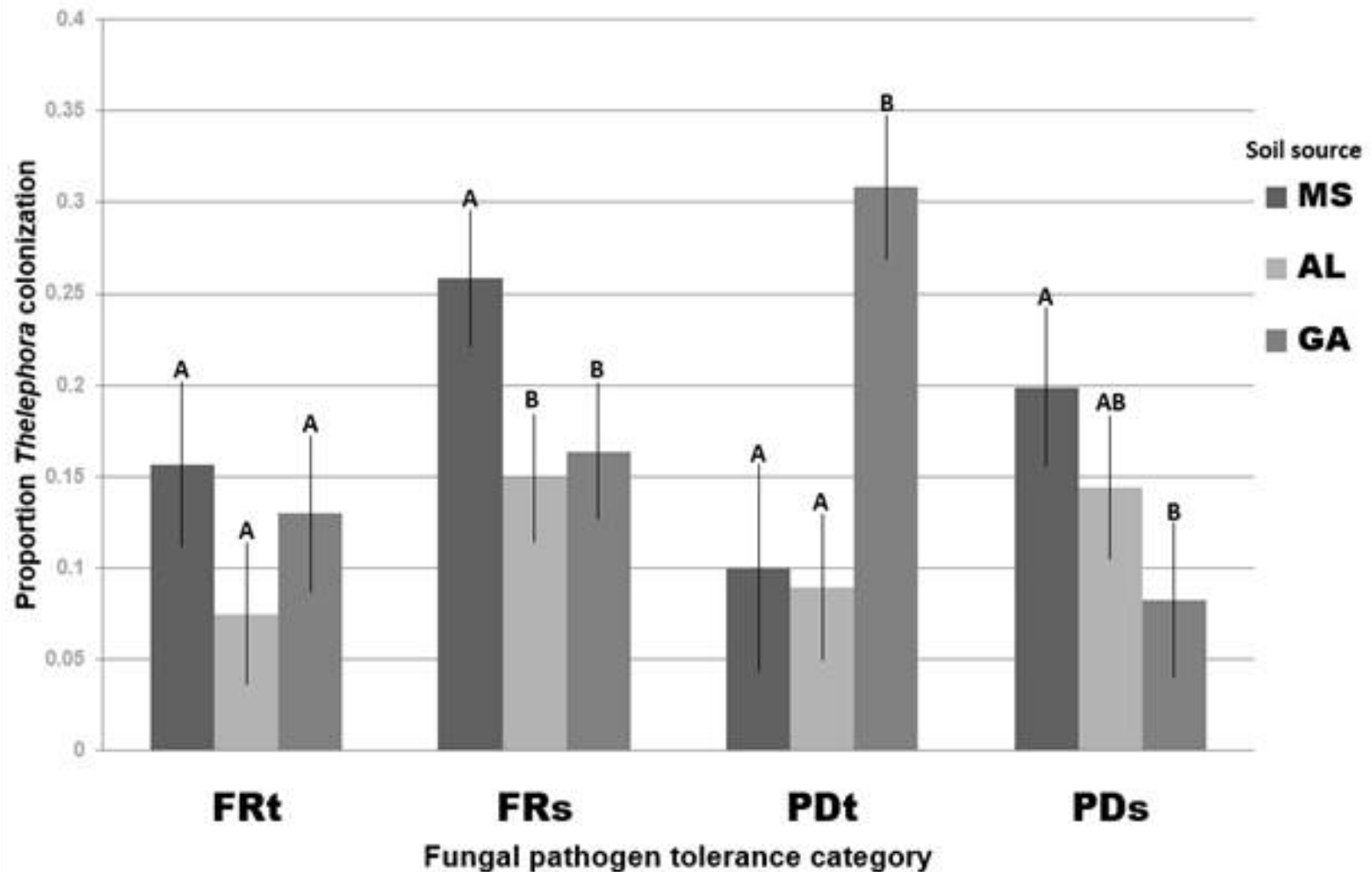
Colonization of *Thelephora* determined by interaction of plant family resistance category and soil inoculation source

($F_{6,536} = 3.691$, $p = 0.00132$)



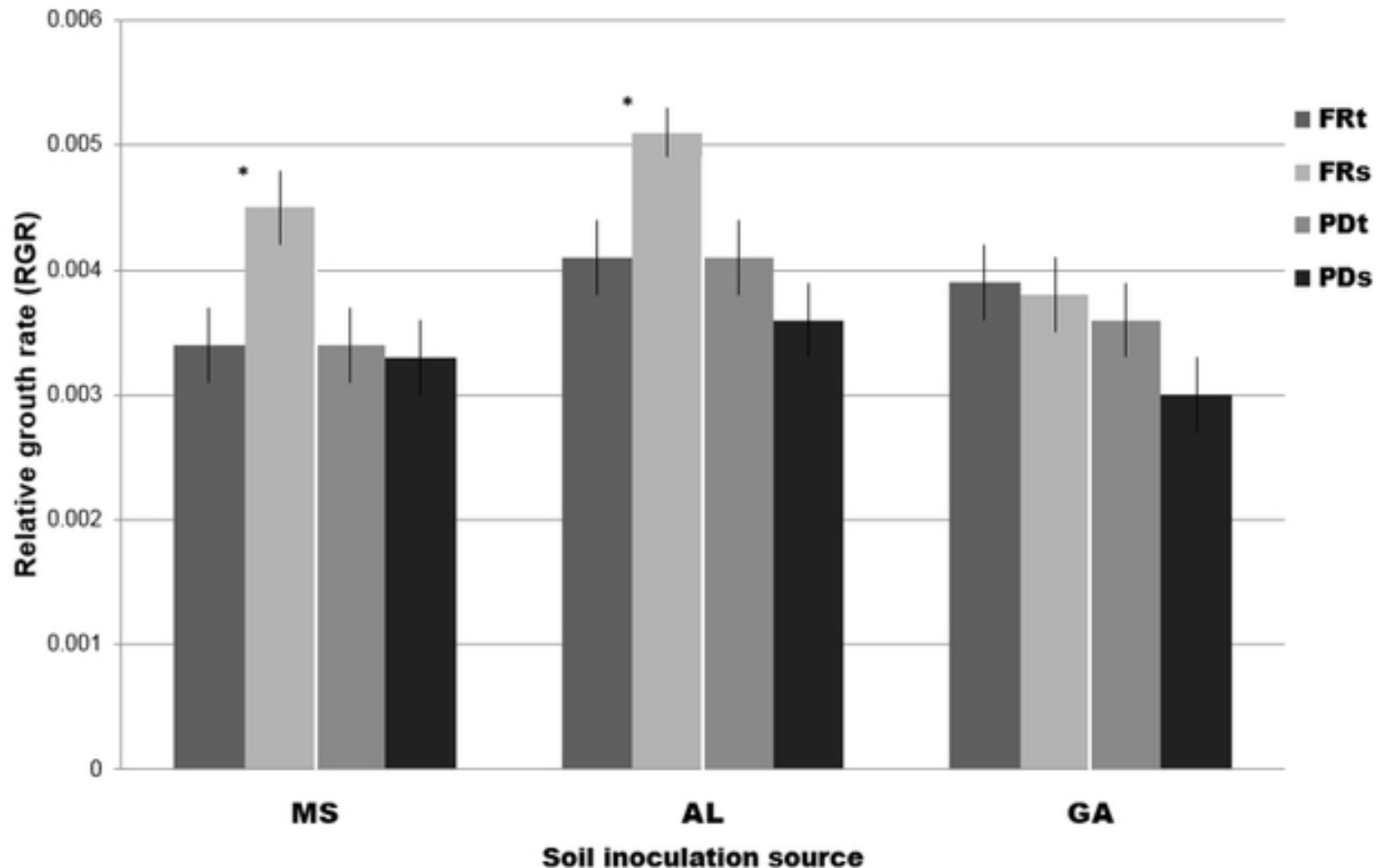
Colonization of *Thelephora* determined by interaction of plant family resistance category and soil inoculation source

($F_{6,536} = 3.691$, $p = 0.00132$)



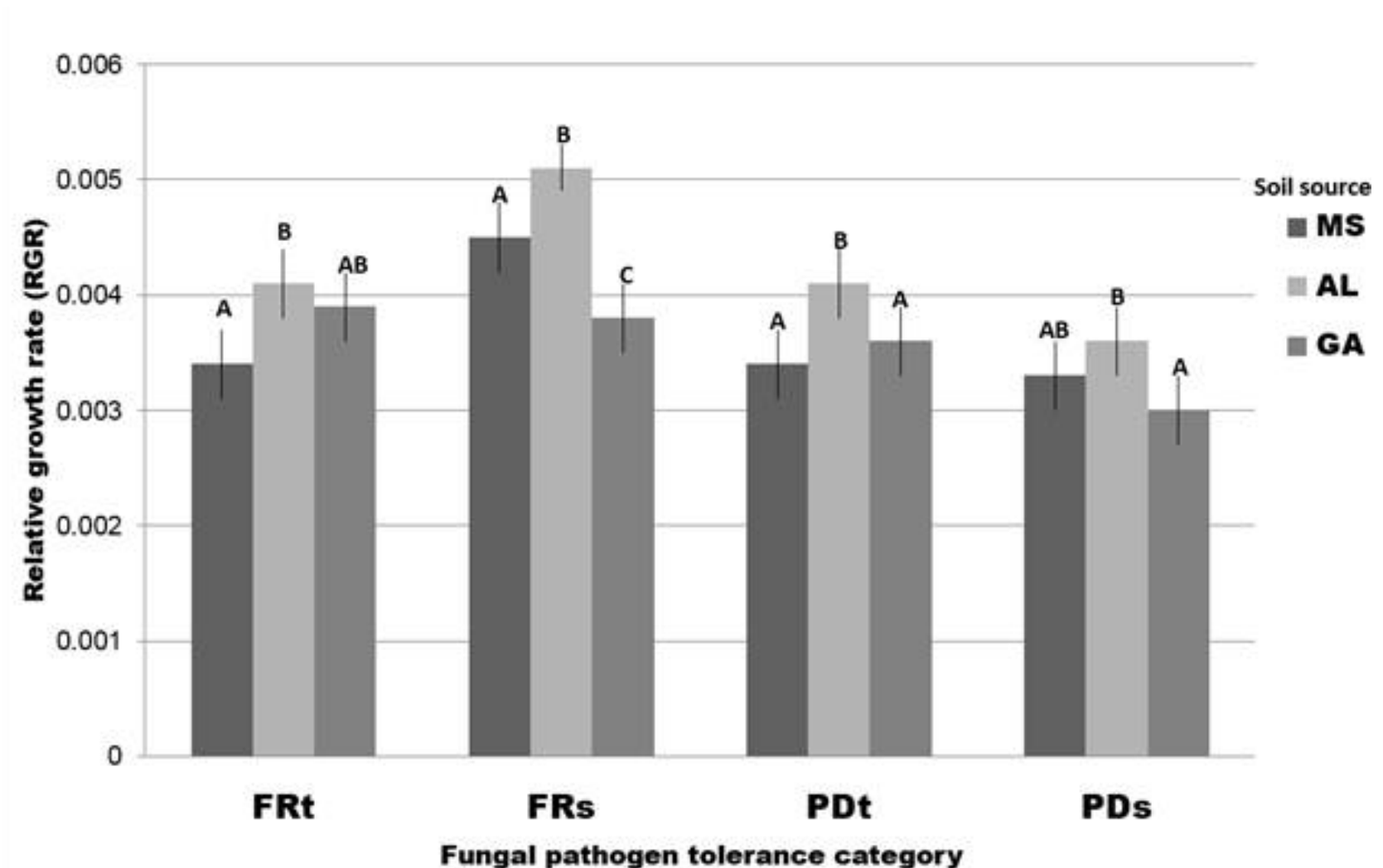
Relative growth rate of seedlings determined by interaction of plant family resistance category and soil inoculation source

($F_{6,532} = 3.342, p = 0.0031$)



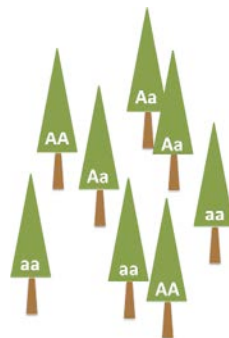
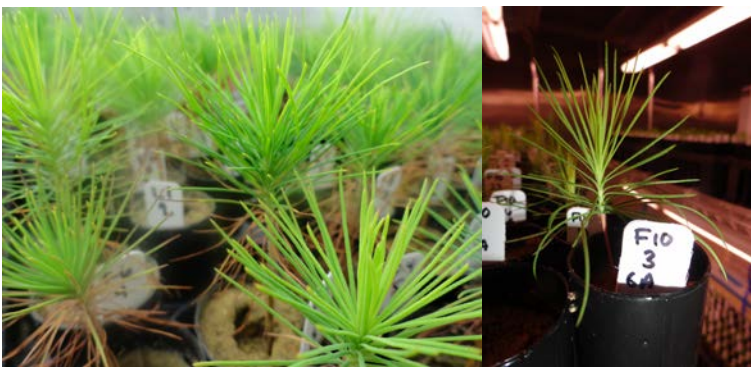
Relative growth rate of seedlings determined by interaction of plant family resistance category and soil inoculation source

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Conclusions

- Evidence of genetic correlations between fungal pathogen tolerance and *Thelephora*
 - Evidence of genetic correlations between fungal pathogen tolerance and relative growth rate
 - outcome of genetic correlations differs among soil environments
 - Loblolly pine interacts with EM species differently
-



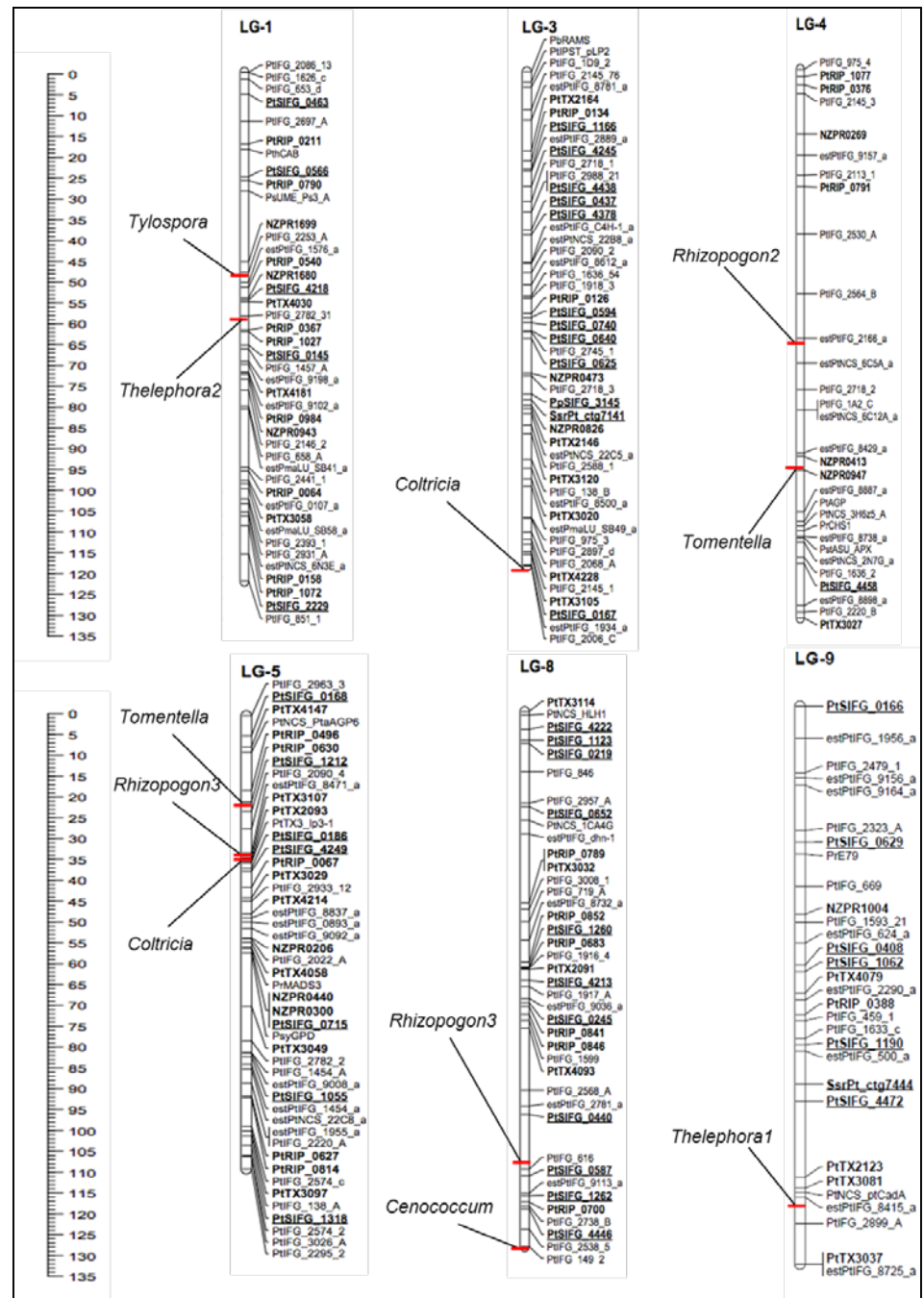
Association mapping of ectomycorrhizal traits in loblolly pine



Ectomycorrhizal traits mapped to loblolly pine genome

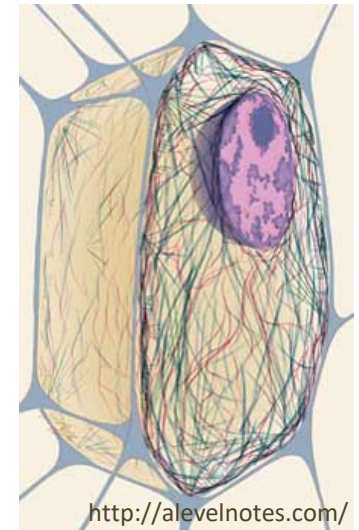
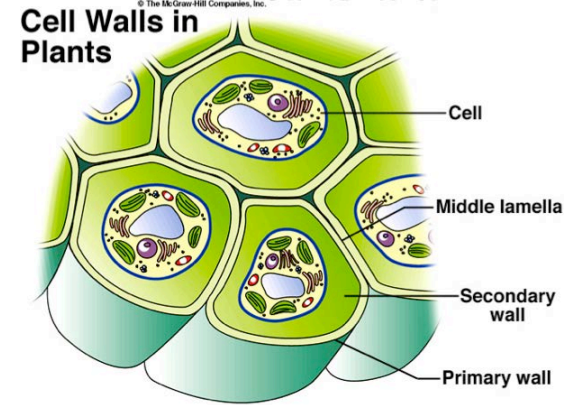
Percentage of phenotypic variance in mycorrhizal traits attributed to marker effects (R^2) ranged from 13% to 55%

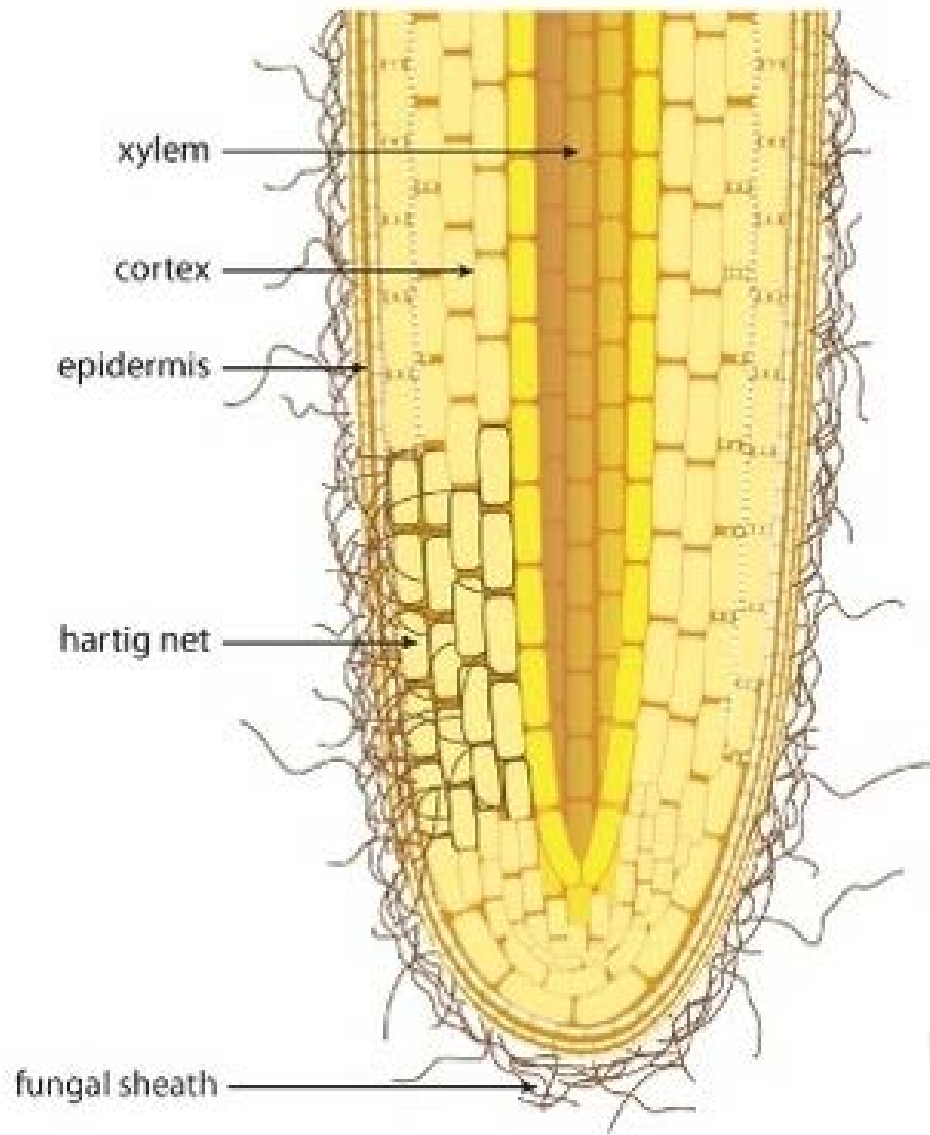
modified from Echt *et al.*, 2011



Host plant cell structural components influence abundance of certain EM fungi

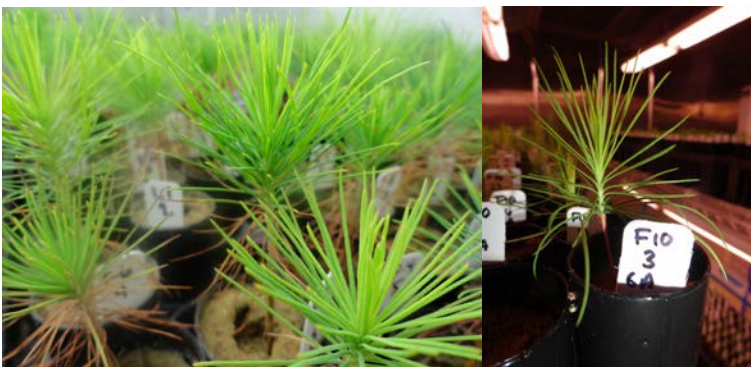
Trait	Annotation	Putative function
Rhizopogon3	Protein domain found in the Ovate family protein (OFP)	In <i>Arabidopsis</i> , <i>OPF4</i> has a role in regulating secondary cell wall formation (Li <i>et al.</i> , 2011)
Rhizopogon3	Glycosyl transferase, family 8 (GT8)	In <i>Arabidopsis</i> , the activity of three members of the GT8 family are involved in the synthesis of xylan (Rennie <i>et al.</i> , 2012), an integral component of plant secondary cell walls
Tomentella	Actin family of proteins	Involved in formation of filaments that are a major component of the cytoskeleton





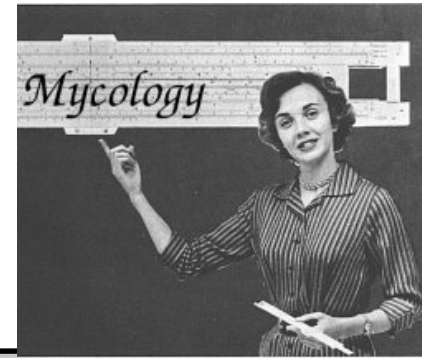
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-
- association analysis suggests fewer genes of large effect
 - mycorrhizal traits associated with host plant cell structural components



Mycorrhizal fungi influence

- **Carbon sequestration**
- **Water stress mitigation**



REPORT

Roots and Associated Fungi Drive Long-Term Carbon Sequestration in Boreal Forest

K. E. Clemmensen^{1,*}, A. Bahr², O. Ovaskainen³, A. Dahlberg^{1,4}, A. Ekblad⁵, H. Wallander², J. Stenlid¹, R. D. Finlay¹, D. A. ...

+ See all authors and affiliations

Science 29 Mar 2013:
Vol. 339, Issue 6127, pp. 1615-1618
DOI: 10.1126/science.1231923

Research review

Mycorrhizal fungi have a potential role in soil carbon storage under elevated CO₂ and nitrogen deposition

K. K. TRESEDER* AND M. F. ALLEN

Center for Conservation Biology, University of California at Riverside, Riverside, CA 92521, USA

Received 9 November 1999; accepted 7 March 2000

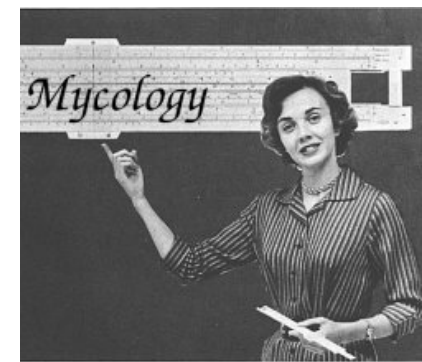
Global Change Biology

PRIMARY RESEARCH ARTICLE | [Free Access](#)

Mycorrhizal fungi enhance plant nutrient acquisition and modulate nitrogen loss with variable water regimes

Timothy M. Bowles✉, Louise E. Jackson, Timothy R. Cavagnaro

First published: 01 September 2017 | <https://doi.org/10.1111/gcb.13884>



The Response Patterns of Arbuscular Mycorrhizal and Ectomycorrhizal Symbionts Under Elevated CO₂: A Meta-Analysis

Yuling Dong^{1,2}, Zhenyu Wang³, Hao Sun^{1,2}, Weichao Yang¹ and Hui Xu^{1*}

¹ Key Laboratory of Pollution Ecology and Environmental Engineering, Institute of Applied Ecology, Chinese Academy of Sciences, Shenyang, China, ² University of Chinese Academy of Sciences, Beijing, China, ³ School of Biological and Chemical Engineering, Liaoning Institute of Science and Technology, Benxi, China

“Our results demonstrated that eCO₂ increased mycorrhizal plants biomass (+26.20%) . . . and mycorrhizal fungal growth (+22.87% in extraradical hyphal length and +21.77% in mycorrhizal fungal biomass)”

Acknowledgements

Jason Hoeksema (Ole Miss)
Lori Eckhardt (Auburn)
C. Dana Nelson (USFS)
Jim Roberds (USFS)
Pedro Jose Martinez Garcia (UC Davis)
Seth Pritchard (C of C)
Allan Strand (C of C)
Chuck Burdine (USFS)



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Awesome Undergrads!

Whitney Sephaus

Sophie Kreutz

Julie Raguel

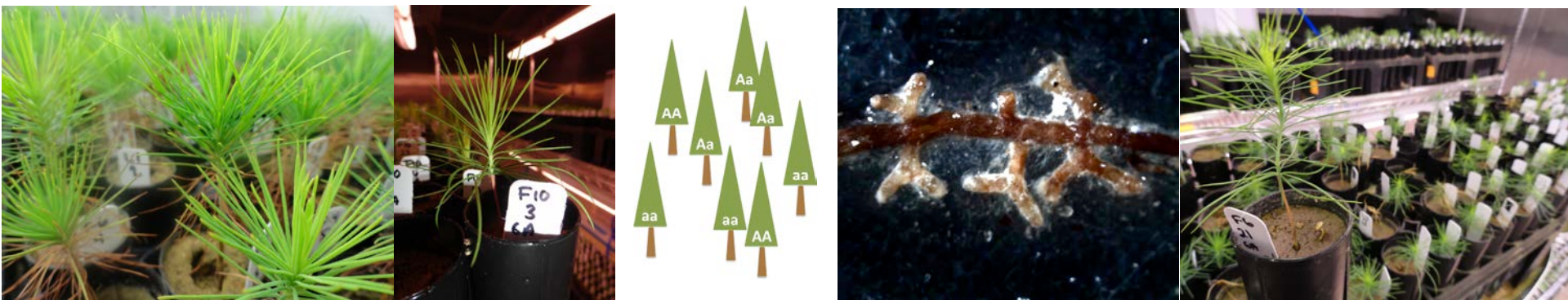


Questions?

Bridget Piculell
piculellb@cofc.edu

Conclusions

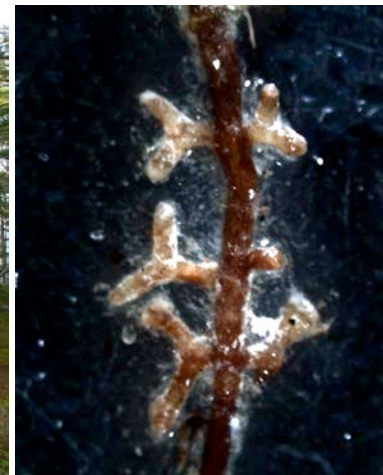
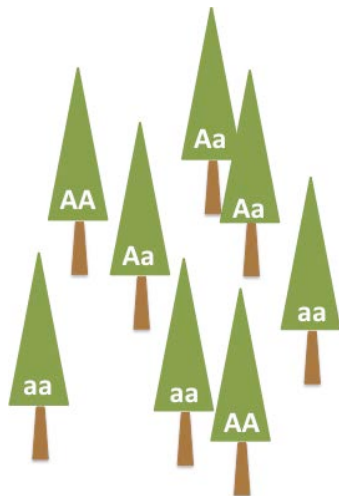
- environmental variation has more influence on mycorrhizal community than genetic variation ***** New analysis needed with corrected pedigree!
 - genetic correlations between mycorrhizal traits and other traits
-
- association analysis suggests fewer genes of large effect
 - shared chromosome regions confirmed by quantitative genetic analysis
 - mycorrhizal traits associated with host plant cell structural components
-



1: Heritability and genetic correlations of above- and belowground traits in loblolly pine

2: Association mapping of ectomycorrhizal traits in loblolly pine

3: Current work at College of Charleston



What happens when roots die?

Fine root senescence is understudied

- Soil carbon pools are much larger than most other carbon pools on the planet
- Fine root turnover represents the most significant mode of carbon flux from plants into these pools, outstripping the role of leaves and other aboveground litter.



What happens when roots die?

Fine root senescence is understudied

- To track fine roots through time and access them for manipulations in the field, we have installed 100 30x30cm plastic “root windows” at a site in coastal SC



What happens when roots die?

Fine root senescence is understudied

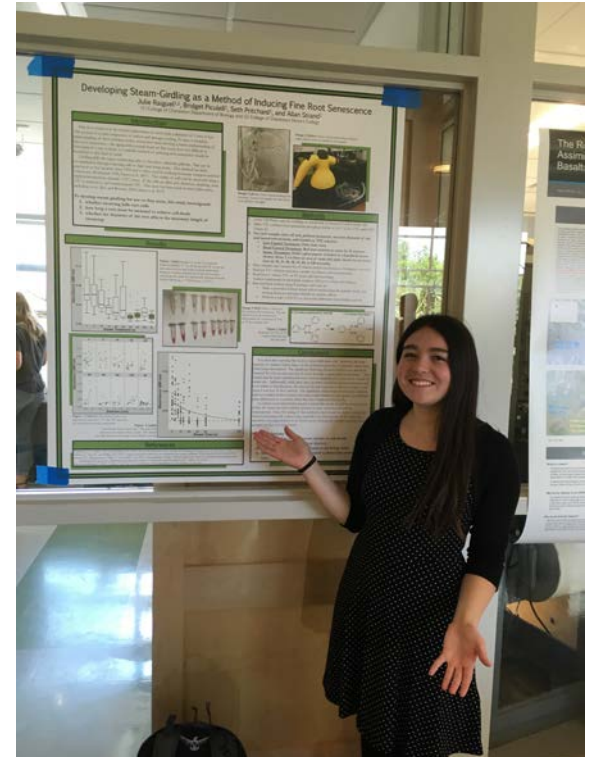
- Toobis (tractable observation of biotic interactions*)



What happens when roots die?

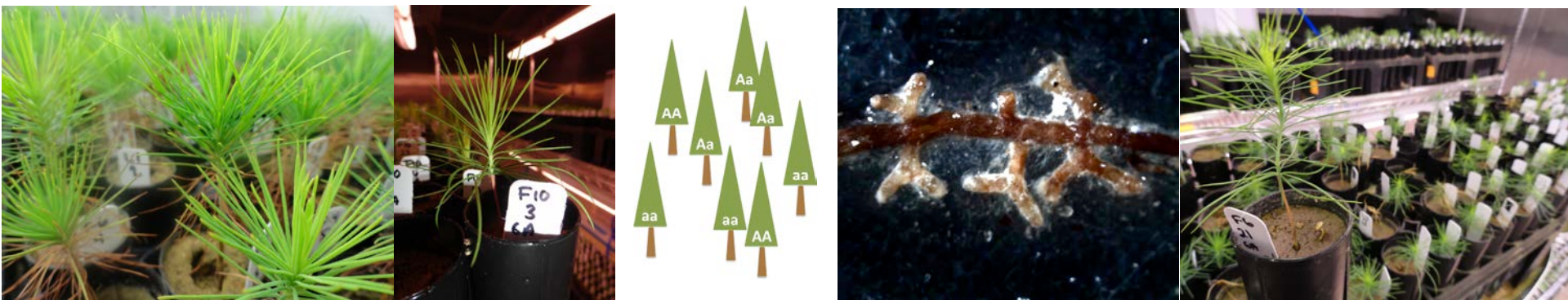
Fine root senescence is understudied

- Steam girdling as a method to induce root senescence



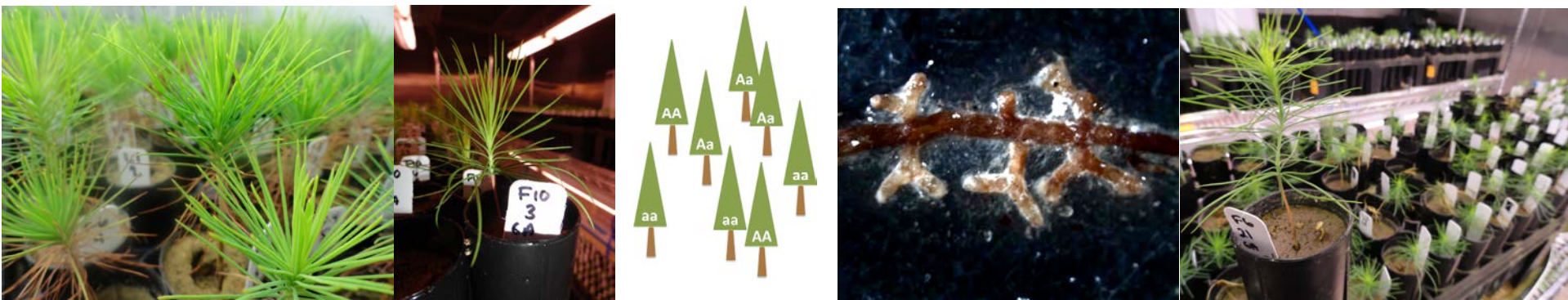
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 - mycorrhizal traits associated with host plant cell structural components
-



Synthesis

- Loblolly pines evolve independently in their interactions with different EM fungi
- Genetic correlations exist both among EM traits and between EM and other traits
- There is variation in the expression of genetic correlations
- Genetic determination of EM traits may be linked to cell structure

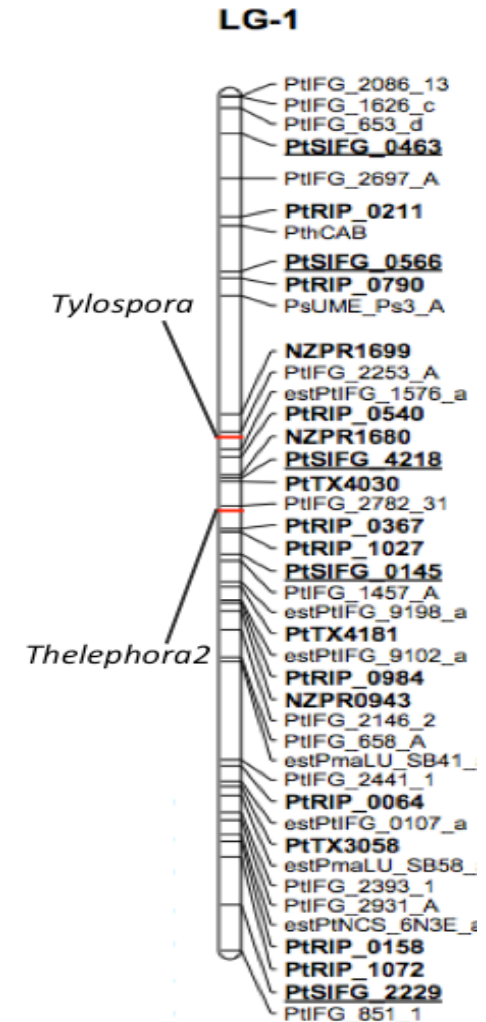


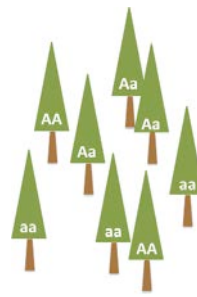


Quantitative genetic analysis

SNP association analysis

Chromosome	trait - trait	Pearson's r	p value
1	Tylospora - Thelephora2	0.27	0.00011

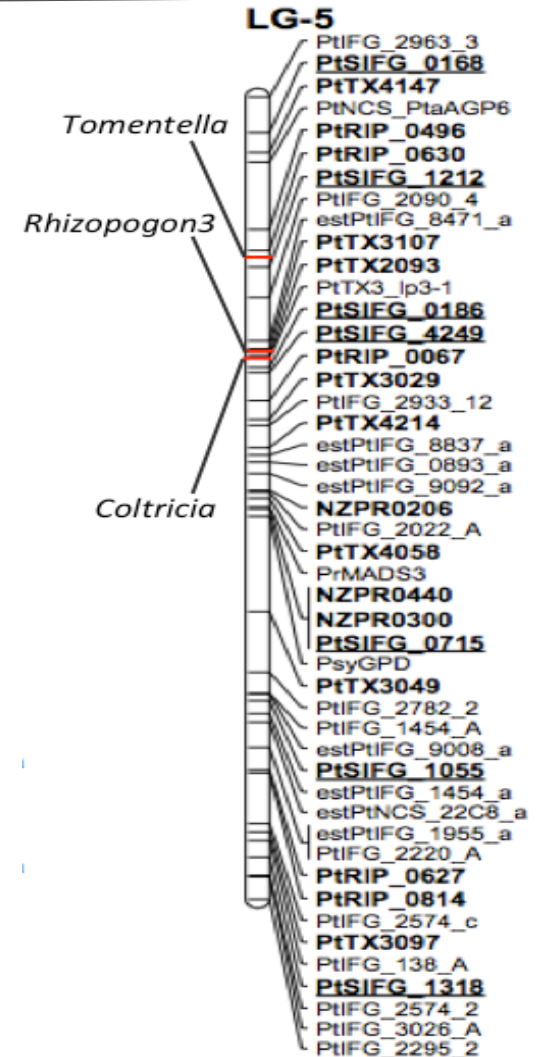


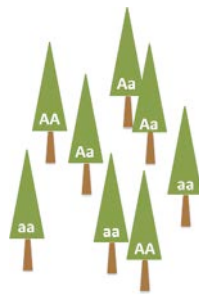


Quantitative genetic analysis

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1	Tylospora - Thelephora2	0.27	0.00011
5	Tomentella - Rhizopogon3	0.34	1.02 E -6
	Coltricia - Rhizopogon3	0.19	0.00011
	Coltricia - Tomentella	0.28	8.5 E -5

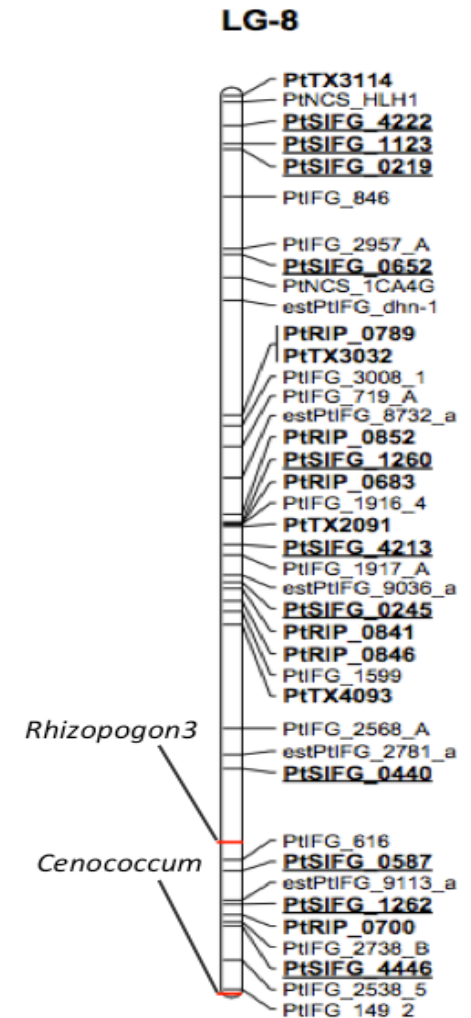


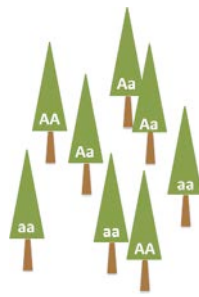


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	Coltricia - Tomentella	0.28	8.5 E -5
8	Cenococcum - Rhizopogon3	-0.17	0.017

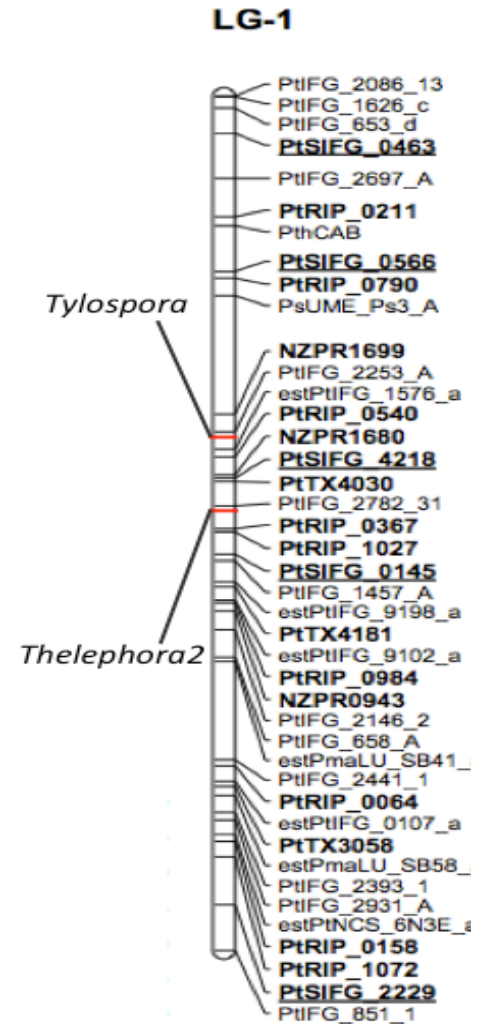


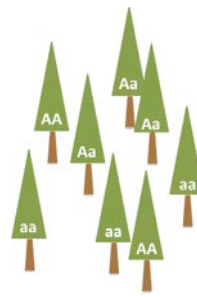


Quantitative genetic analysis

SNP association analysis

Chromosome	trait - trait	Pearson's r	p value
1	Tylospora - Thelephora2	0.274	0.00011

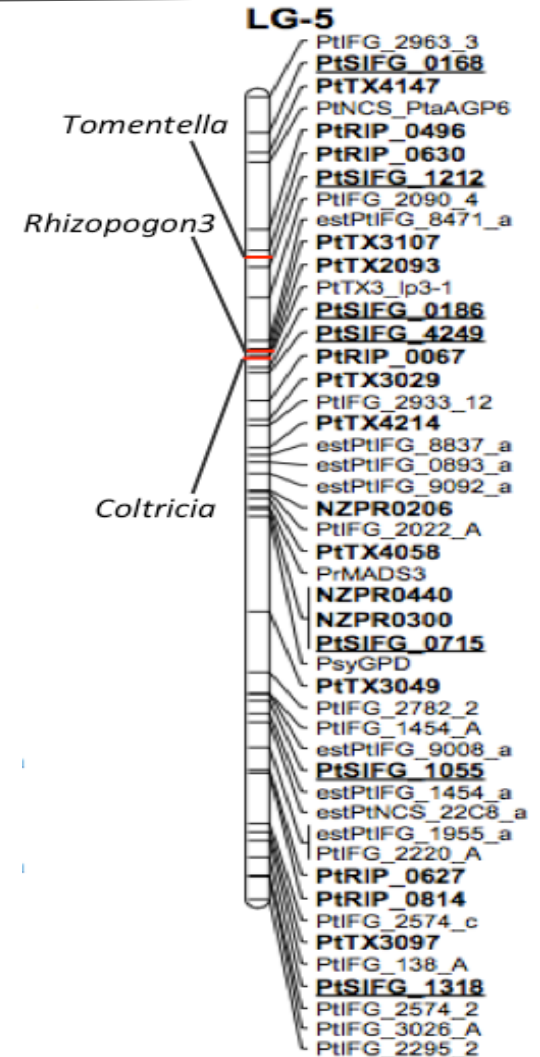


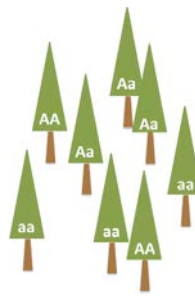


Quantitative genetic analysis

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1	Tylospora - Thelephora2	0.274	0.00011
5	Tomentella - Rhizopogon3	0.34245	1.02 E -6
	Coltricia - Rhizopogon3	0.1859	0.00011
	Coltricia - Tomentella	0.27836	8.5 E -5

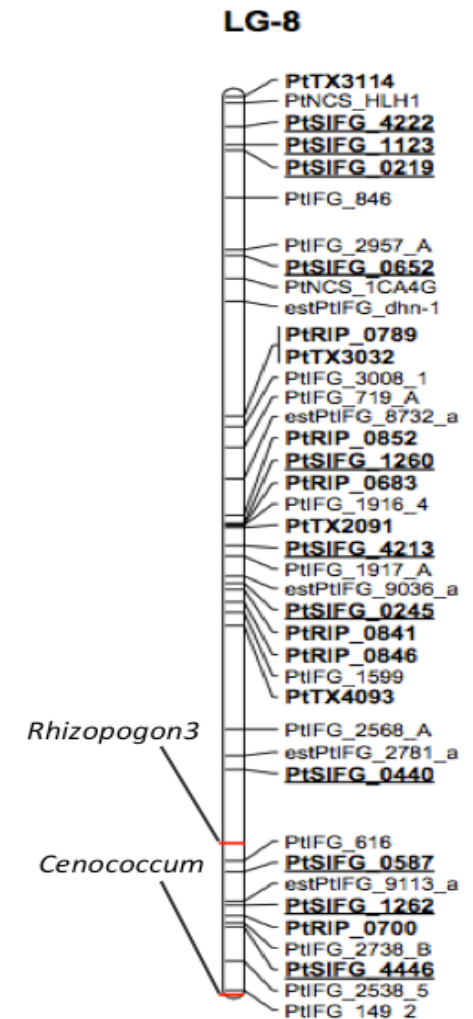




Quantitative genetic analysis

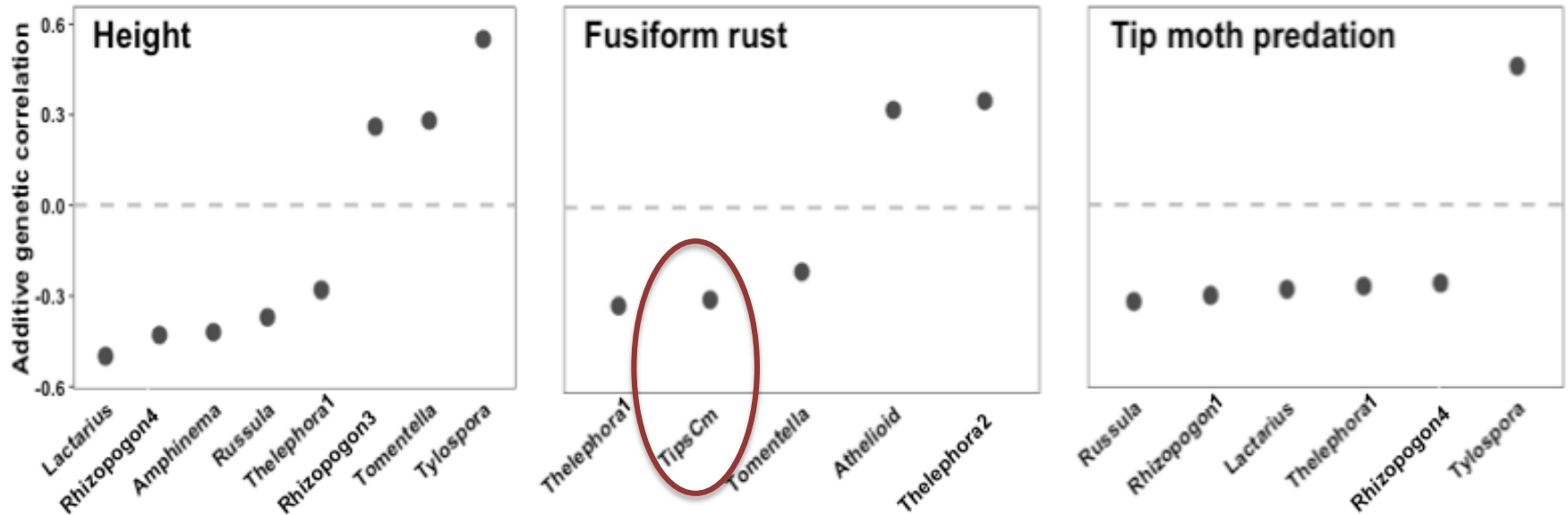
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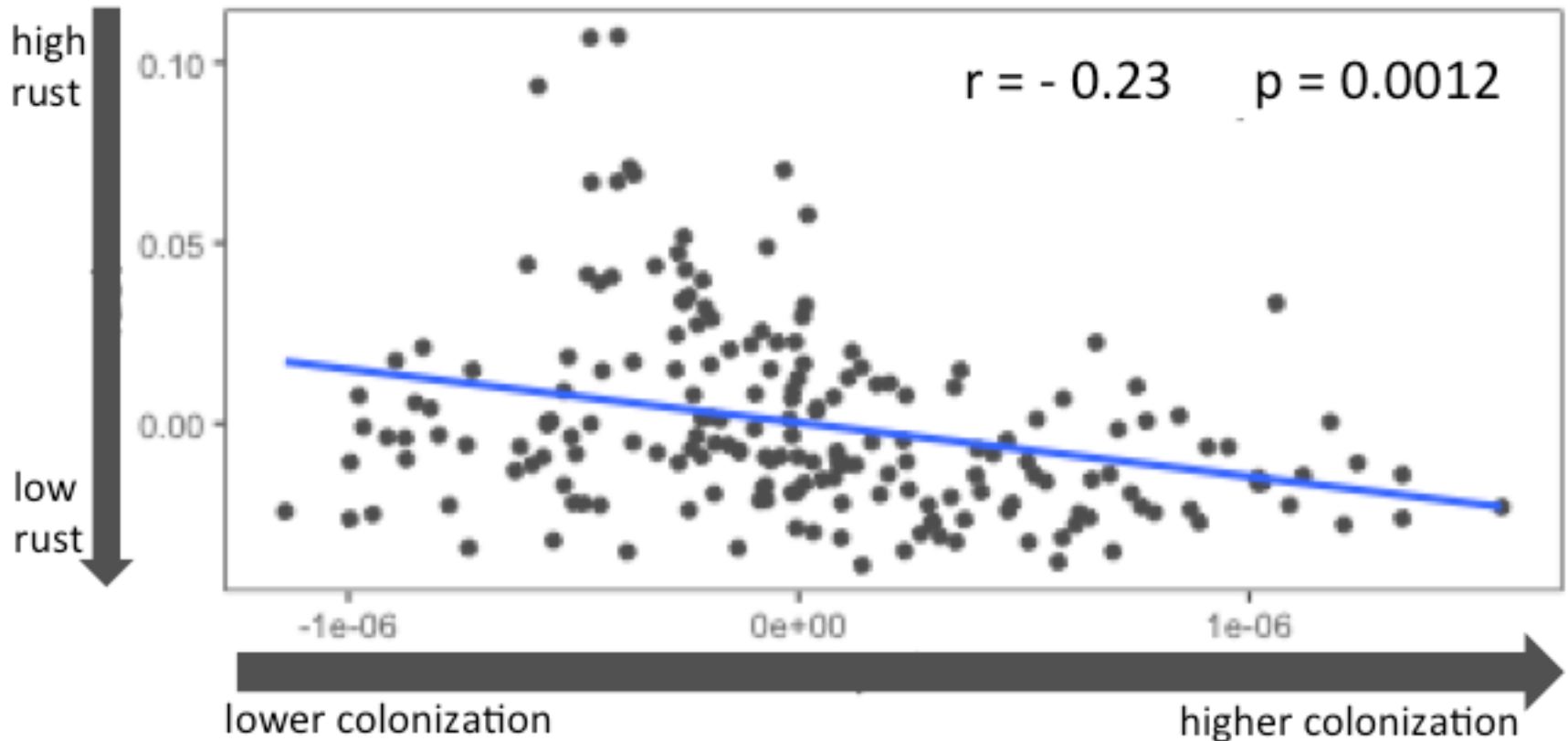


Genetic correlations between traits in loblolly pine

Pearson's r shown for all significant correlations ($p < 0.05$)

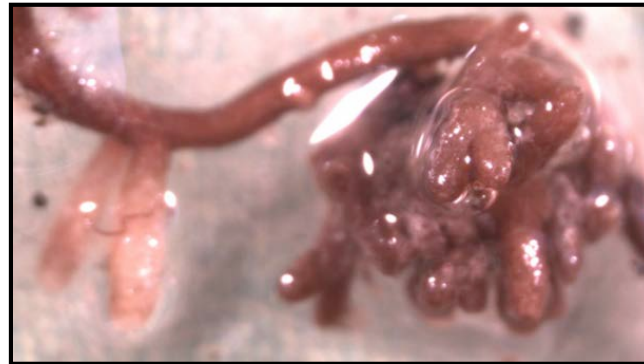


Genetic correlations between fusiform rust infection and mycorrhizal colonization in loblolly pine

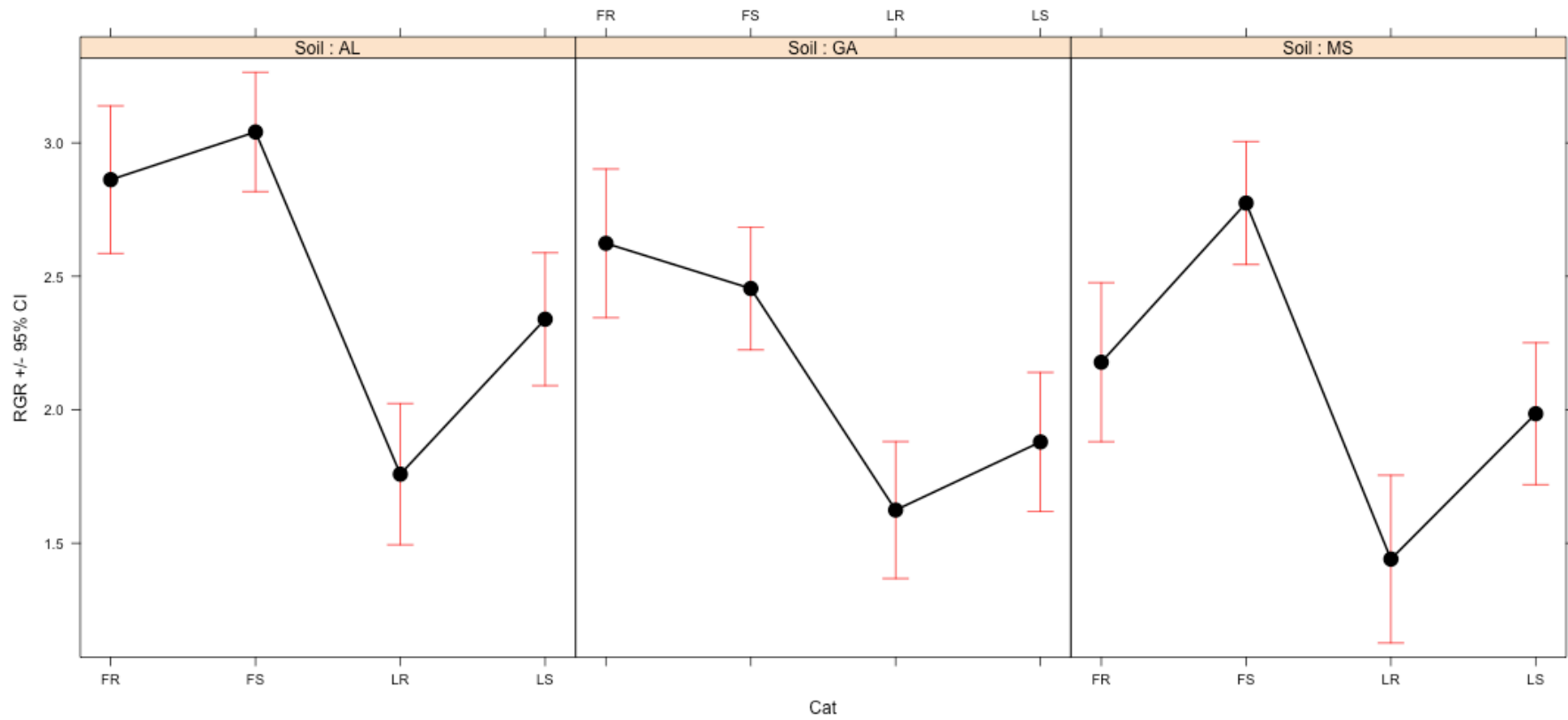


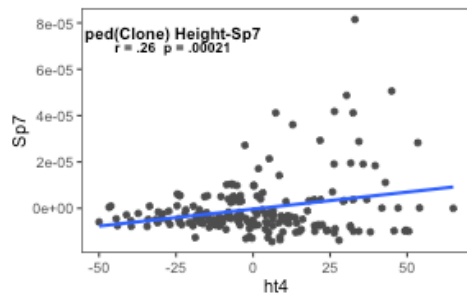
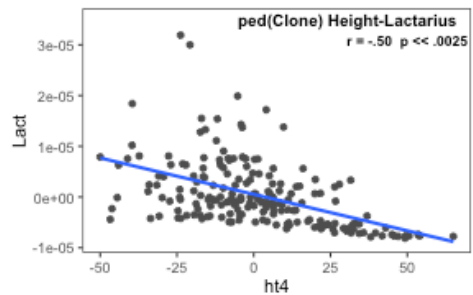
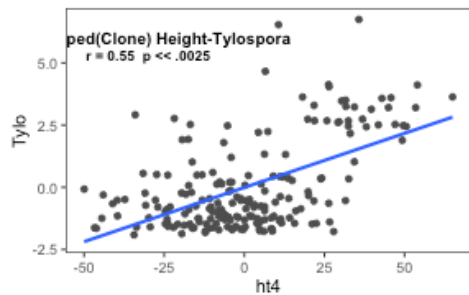
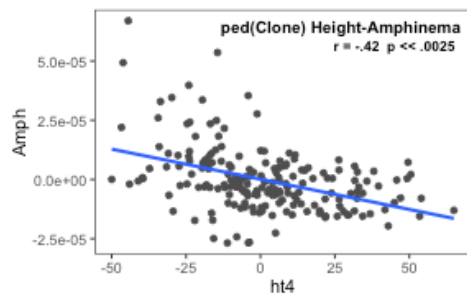
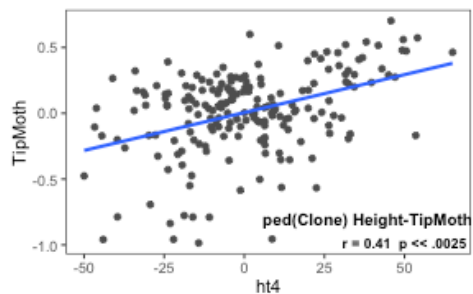
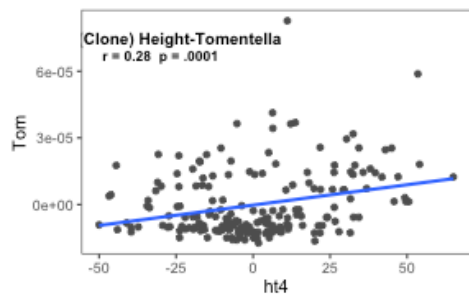
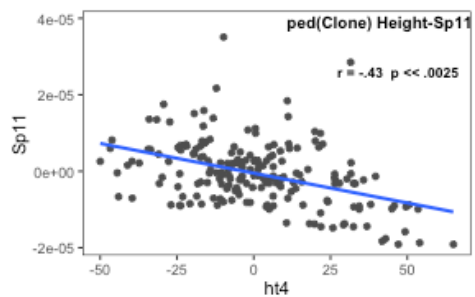
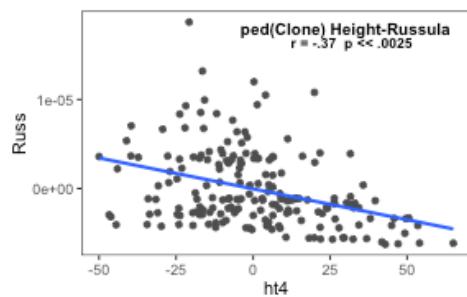
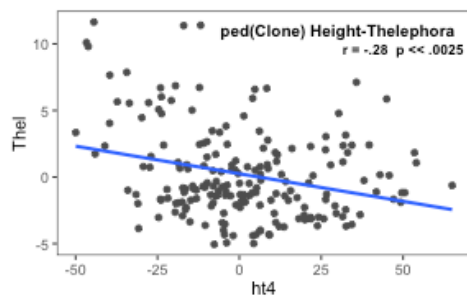
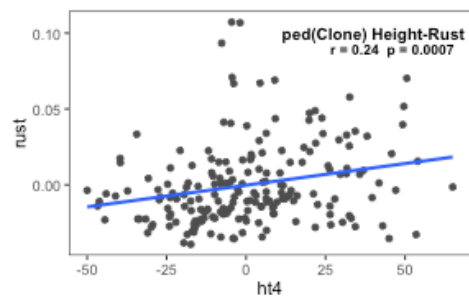
Morphotyping and fungal identification

Root tips colonized by ectomycorrhizal fungi grouped by similar phenotypic characteristics (color, texture, emanating hyphae)



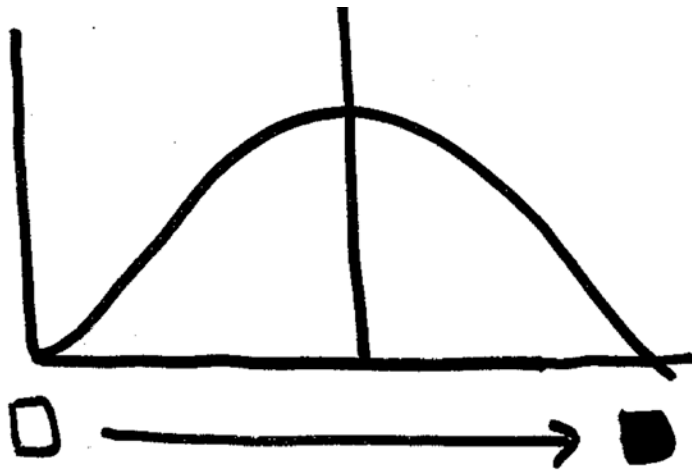
Cat*Soil effect plot



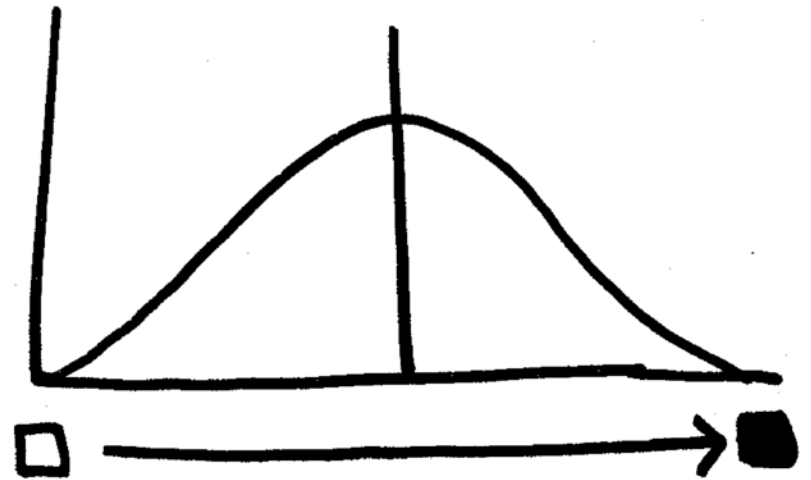




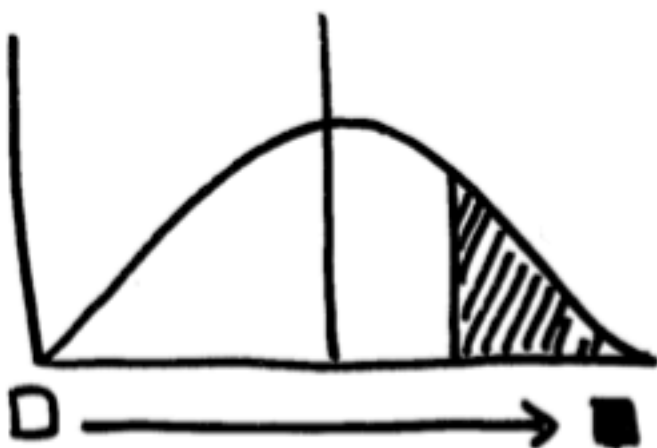
$$V_P = V_A + V_I + V_E$$



$$V_P = V_A + V_D + V_I + V_E$$



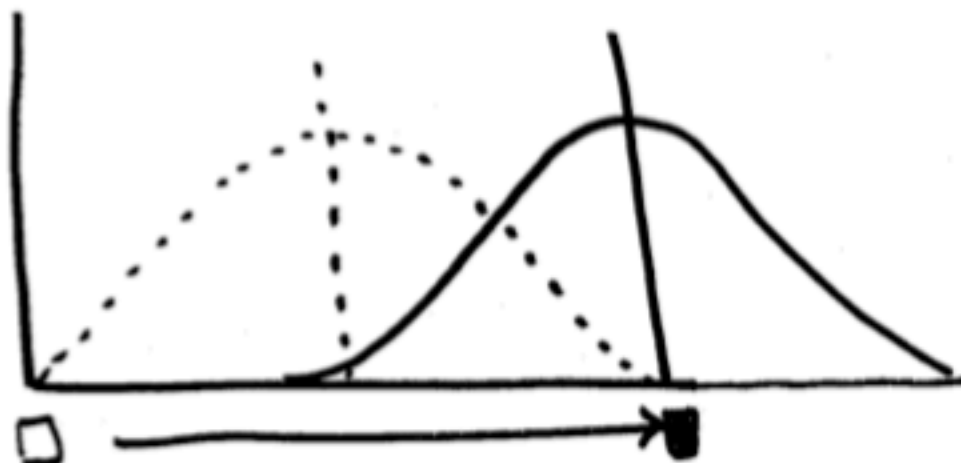
$$V_P = V_A + V_I + V_E$$



$$V_P = V_A + V_D + V_I + V_E$$



NEXT GENERATION





$$V_G = V_A + V_D + V_I$$

$$V_P = V_A + V_D + V_I + V_E$$

V_D -> interaction among alleles at a single locus

V_I -> interaction among genes at different loci (epistasis)

V_A -> represents the cumulative effect of individual loci;
the overall mean is equal to the summed contribution of
these loci

- **resemblance between relatives is caused primarily by additive variation**
- **most important for sexually reproducing species**

$$V_P = V_G + V_E$$

$$V_P = V_A + V_D + V_I + V_E$$

Heritability

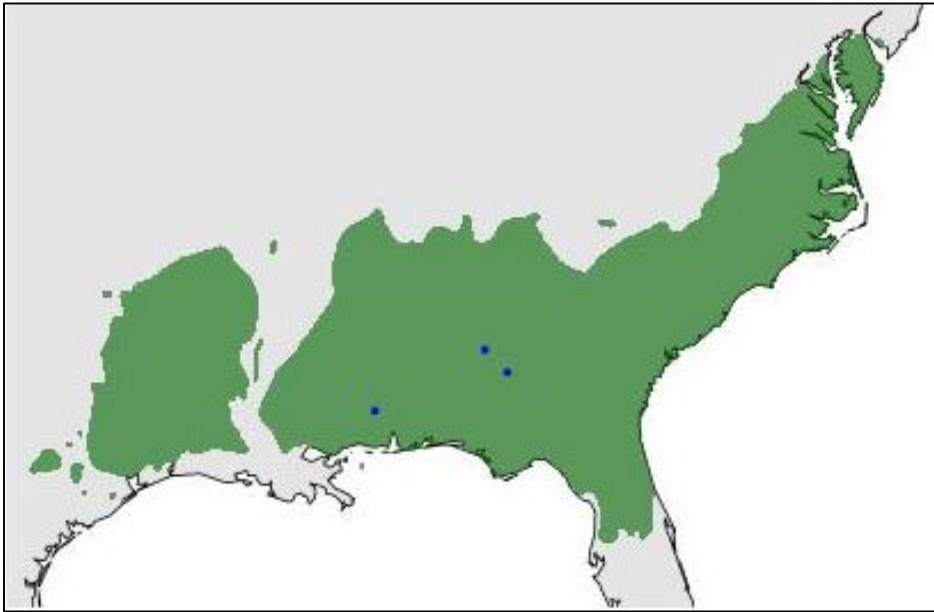
$$H^2 = V_G / V_P \quad (\text{broad sense})$$

$$h^2 = V_A / V_P \quad (\text{narrow sense})$$

Calculating narrow-sense heritability is important for predicting how a trait will respond to selection

Estimating additive genetic variation in loblolly pine

Loblolly Pine, *Pinus taeda*



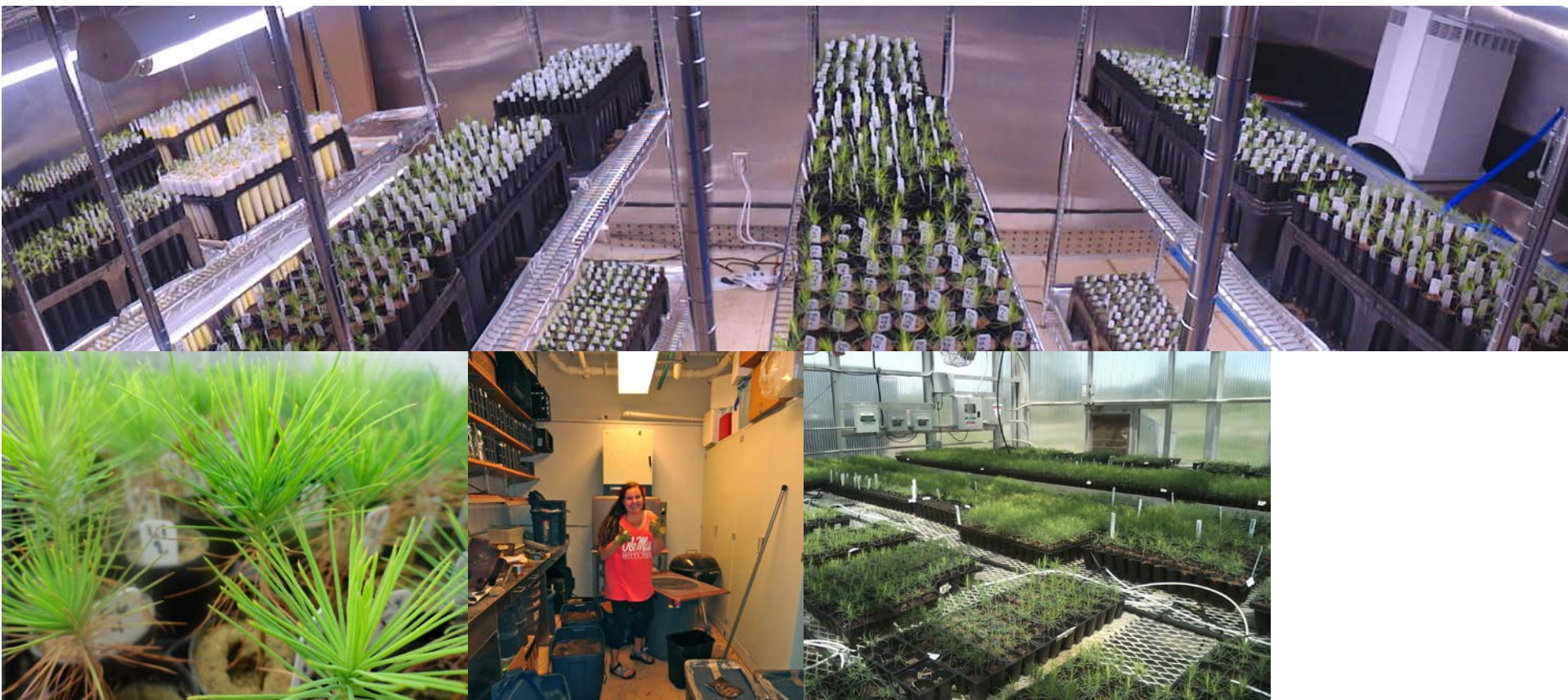


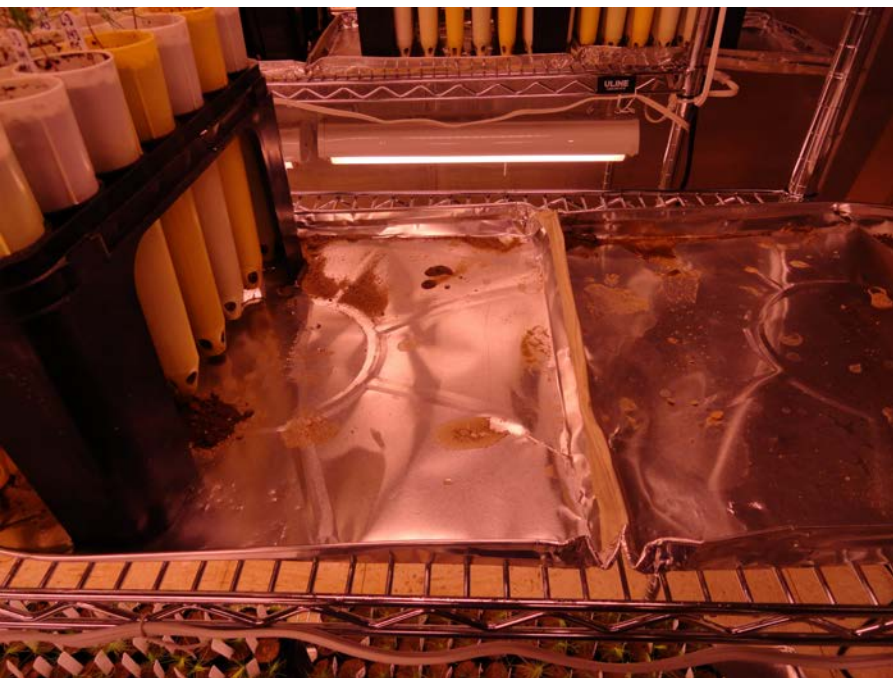


1) 30.615068, -89.059742, 2) 30.615105, -89.058320, 3) 30.614052, -89.059983, 4) 30.614108, -89.058599, 5) 30.615354, -89.058229, 6) 30.614858, -89.057139, 7) 30.614726, -89.058235, 8) 30.614593, -89.057934, 9) 30.614368, -89.057131, 10) 30.614241, -89.056849, 11) 30.614057, -89.057929, 12) 30.613595, -89.056867

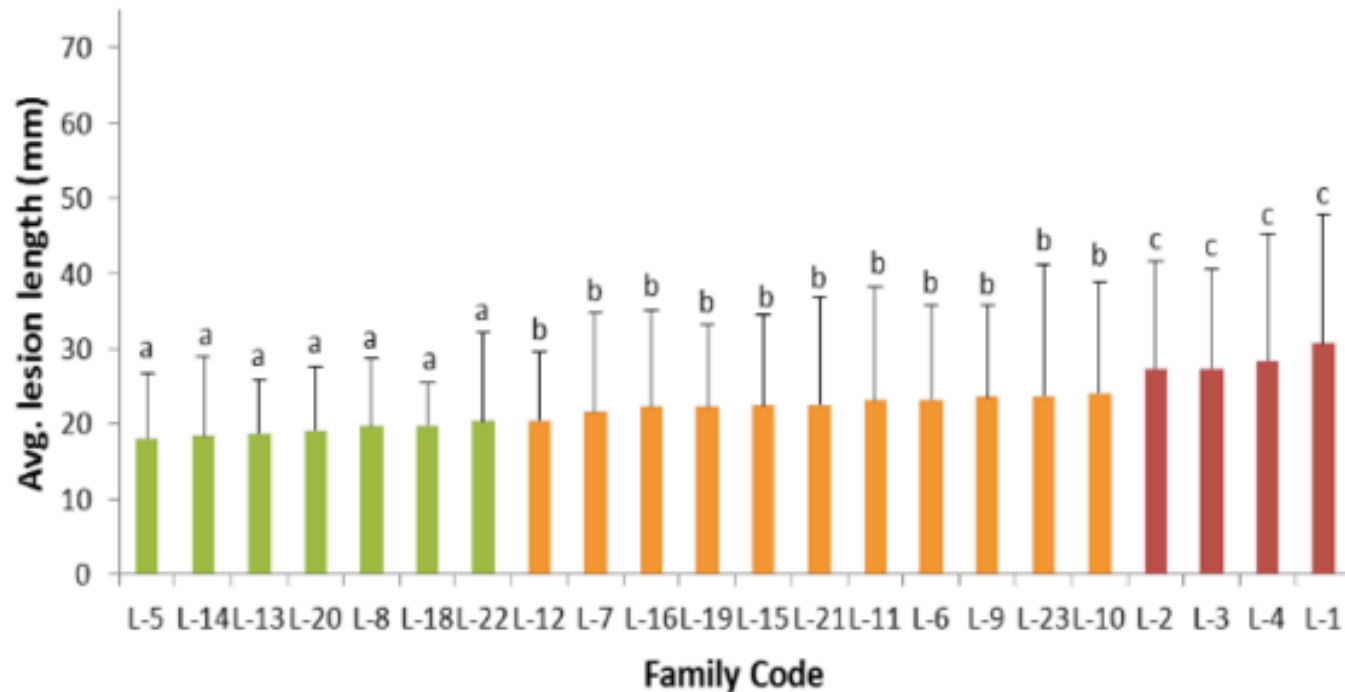
Chapter 1: Genetically determined pathogen resistance interacts with geographic variation to influence mycorrhizal community

Bridget J. Piculell, Lori G. Eckhardt, and Jason D. Hoeksema





Overall Family Ranking



1: Heritability and genetic correlations of above and below ground traits in loblolly pine

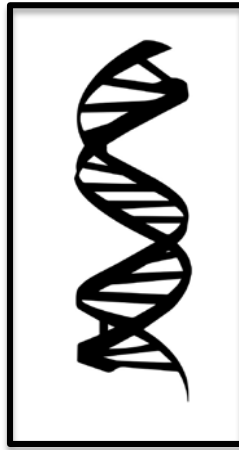


Partitioning variance between genetic and environmental effects



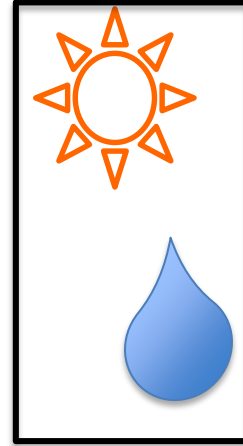
phenotype

=



genotype

+



environment

Partitioning variance between genetic and environmental effects

Phenotype = Genotype + Environment

$$P = G + E \text{ (individual)}$$

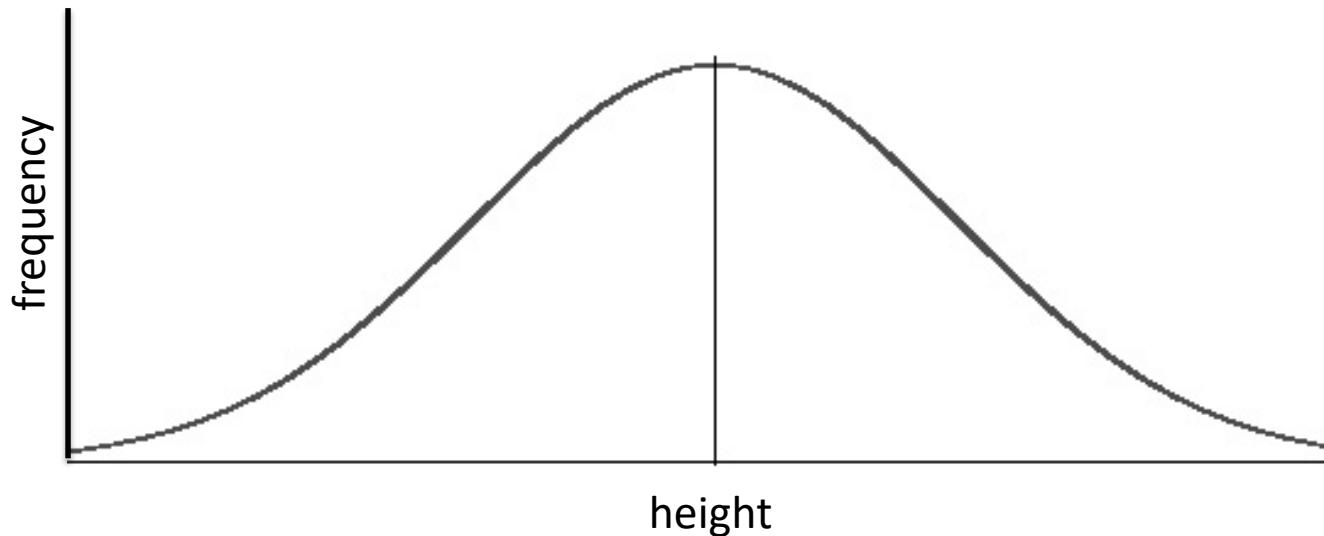


Partitioning variance between genetic and environmental effects

Phenotype = Genotype + Environment

$$P = G + E \quad (\text{individual})$$

$$V_P = V_G + V_E \quad (\text{population})$$



Partitioning variance between genetic and environmental effects

$$V_P = V_G + V_E$$

$$V_P = \overset{\text{additive}}{\left[\underset{\text{A}}{V_A} + \underset{\text{I}}{V_I} \right]} + \overset{\text{non additive}}{V_E}$$

V_A represents the cumulative effect of individual loci

V_I interaction among alleles at a single locus
interaction among genes at different loci

Partitioning variance between genetic and environmental effects

$$V_P = V_G + V_E$$

$$V_P = \overset{\text{additive}}{V_A} + \overset{\text{non additive}}{V_I} + V_E$$

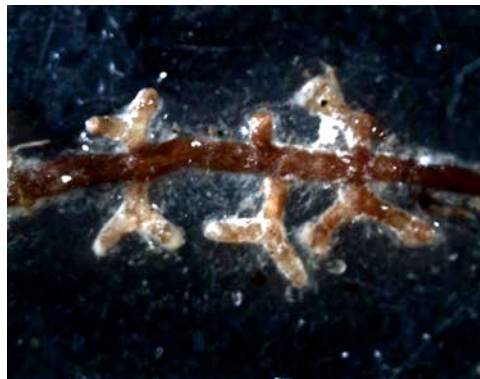
- narrow-sense heritability, $h^2 = V_A / V_P$
- broad-sense heritability, $H^2 = V_G / V_P$
- additive genetic correlations



Heritability and genetic correlations of above and below ground traits in loblolly pine

Q1: How much of the phenotypic variation in a suite of loblolly pine traits is genetically determined?

Q2: Are there additive genetic correlations between mycorrhizal traits of loblolly pine and other traits?



Harrison Experimental Forest (HEF), Southern Institute of Forest Genetics (USDA), Saucier MS



Loblolly pine pedigreed population

Sampling scheme

480 trees



160 genotypes
(x3)



30 families



Phenotypic traits measured

Fungal colonization
(tips/cm)

Fusiform rust
infection

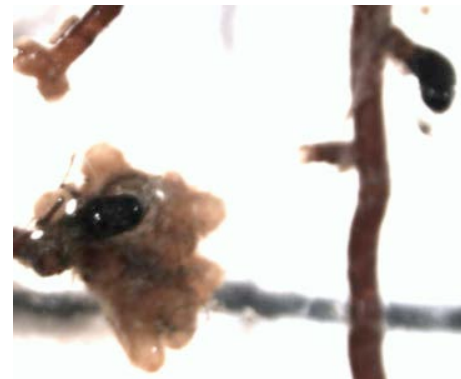
Pine tip moth

Height

Heritability and genetic correlations of above and below ground traits in loblolly pine

Q1: How much of the phenotypic variation in a suite of loblolly pine traits is genetically determined?

Q2: Are there additive genetic correlations between mycorrhizal traits of loblolly pine and other traits?



Heritability estimates for selective traits of loblolly pine

	H^2	h^2
Height	0.179	0.179
Rust	0.0292	0.0292
Tip Moth	0.0791	0.0297

Broad sense heritability, $H^2 = V_A + V_I / V_P$

Narrow sense heritability, $h^2 = V_A / V_P$

Heritability estimates for mycorrhizal traits of loblolly pine

	H^2	h^2
Athelioid	0.0119	< 0.001
Tylospora	0.0143	0.0132
Cenococcum	0.00378	0.00378
Thelephora	0.0197	0.0197
Thelephora2	0.109	< 0.001
Rhizopogon	< 0.001	< 0.001
Tomentella	< 0.001	< 0.001
Lactarius	< 0.001	< 0.001
Russula	< 0.001	< 0.001
Amphinema	< 0.001	< 0.001
Coltricia	< 0.001	< 0.001
Rhizopogon2	< 0.001	< 0.001
Rhizopogon3	< 0.001	< 0.001
Rhizopogon4	< 0.001	< 0.001
Tot. tips/cm root	< 0.001	< 0.001

Broad sense heritability, $H^2 = V_A + V_I / V_P$

Narrow sense heritability, $h^2 = V_A / V_P$

Heritability and genetic correlations of above and below ground traits in loblolly pine

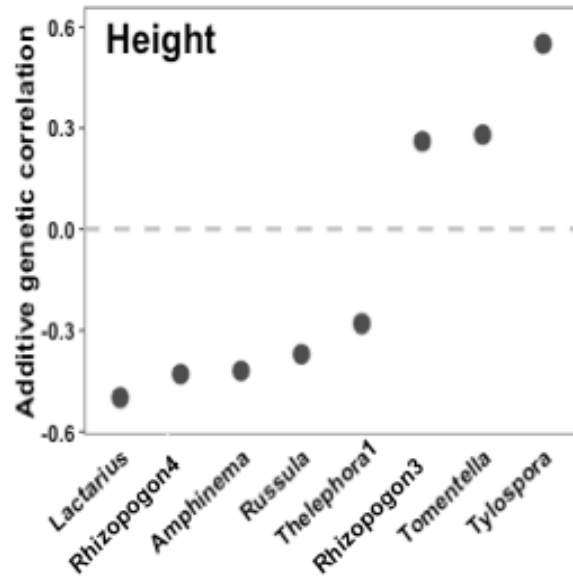
Q1: How much of the phenotypic variation in a suite of loblolly pine traits is genetically determined and how much of the genetic variation is due to either additive genetic variation or interactive genetic effects such as dominance and epistasis?

Q2: Are there additive genetic correlations between mycorrhizal traits of loblolly pine and other traits?



Genetic correlations between traits in loblolly pine

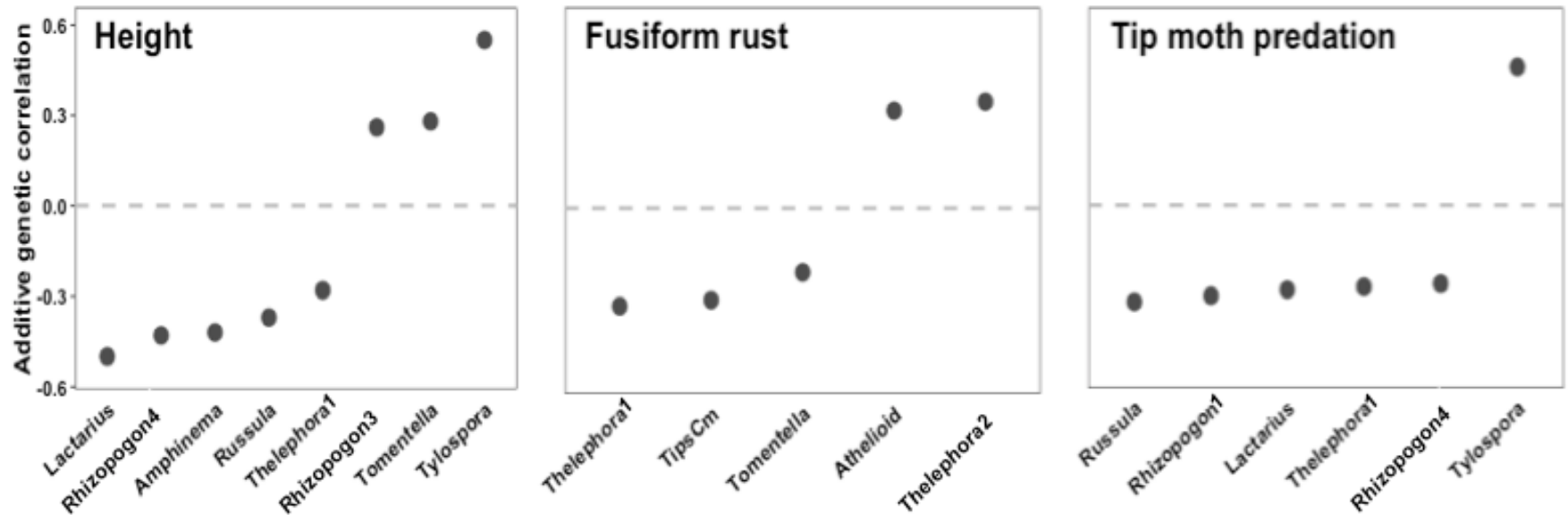
Pearson's r shown for all significant correlations ($p < 0.05$)





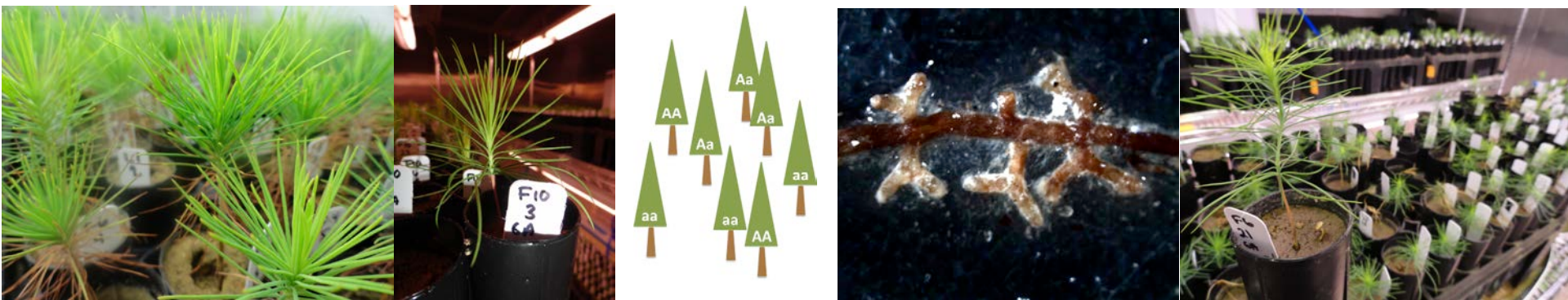
Genetic correlations between traits in loblolly pine

Pearson's r shown for all significant correlations ($p < 0.05$)



Conclusions

- environmental variation has more influence on mycorrhizal community than genetic variation
- genetic correlations between mycorrhizal traits and other traits



2: Association mapping of ectomycorrhizal traits in loblolly pine



Harrison Experimental Forest (HEF), Southern Institute of Forest Genetics (USDA), Saucier MS





SNP Associations with loblolly traits

Illumina Infinium SNP chip

2923 SNPs (1420 informative)

160 genotypes (152 successful)

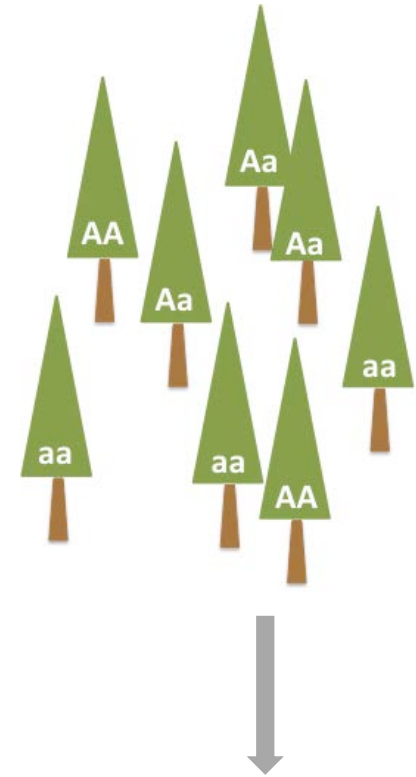
20 loblolly pine traits:

Height

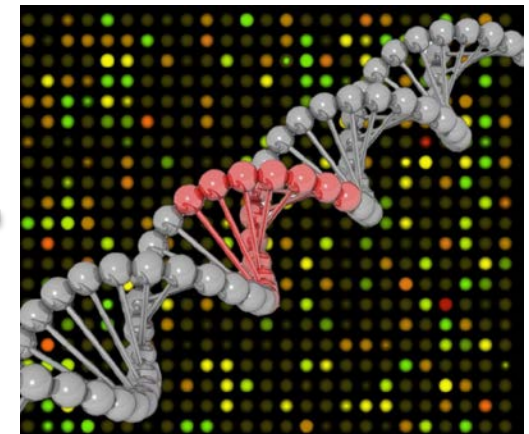
Rust

Tip moth

Fungal colonization traits



74517	A:G	A:G	G:G	A:A	A:A	G:G	C:C	A:G	G:G
74519	A:A	G:G	A:G	A:C	A:C	A:G	C:C	A:G	G:G
74521	A:A	G:G	A:G	A:C	A:C	A:G	A:C	A:G	A:G
74522	A:A	G:G	A:G	A:C	A:C	G:G	C:C	A:A	G:G
74612	A:G	A:G	G:G	A:A	A:C	A:G	C:C	A:A	G:G
74618	A:A	G:G	G:G	A:A	A:C	A:G	C:C	A:A	G:G
74623	A:G	A:G	G:G	A:A	A:A	G:G	C:C	A:A	G:G
74628	A:A	G:G	G:G	A:A	A:C	A:G	C:C	A:A	G:G
74711	A:A	G:G	G:G	A:A	A:C	G:G	C:C	A:G	G:G
74716	A:G	A:G	G:G	A:A	A:C	A:G	C:C	A:A	G:G



Association mapping of ectomycorrhizal traits in loblolly pine

Q1: What can we say about the number and effect of genes involved in mycorrhizal traits in loblolly pine?

Q2: Can we determine a genetic basis for the correlations detected in the quantitative genetics study?

Q3: Can we ascribe putative function to any of the relevant SNPs?



Association mapping of ectomycorrhizal traits in loblolly pine

Q1: What can we say about the number and effect of genes involved in mycorrhizal traits in loblolly pine?

Q2: Can we determine a genetic basis for the correlations detected in the quantitative genetics study?

Q3: Can we ascribe putative function to any of the relevant SNPs?





SNP Associations with loblolly traits

Rhizopogon3 9

Coltricia 7

Cenococcum 4

Rhizopogon4 3

Tomentella 3

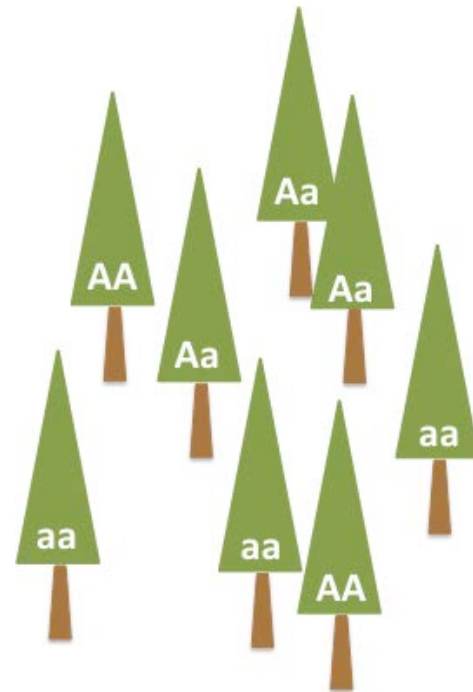
Rhizopogon2 2

Thelephora2 2

Tylospora 2

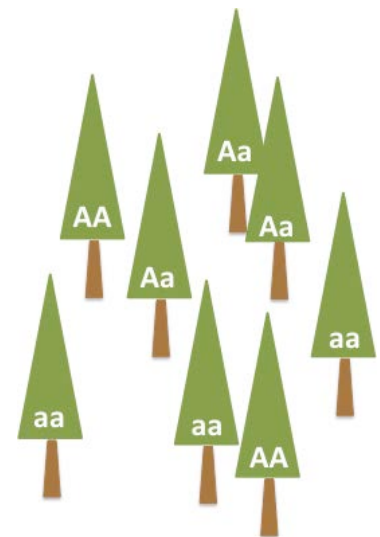
Rust 1

33 significant associations
at $p \leq 3.5 \times 10^{-5}$



Many genes of small effect or few genes of large effect?

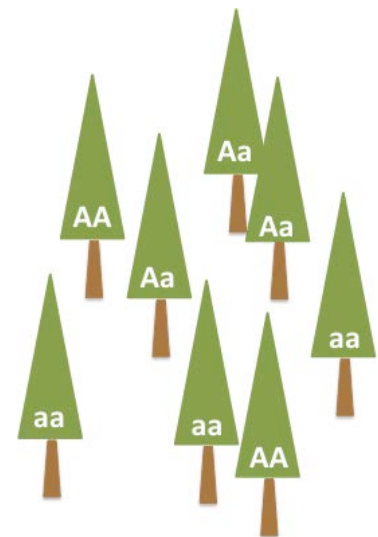
The number of loci controlling a trait can affect the degree and frequency of local adaptation (Nuismer et al., 2007; Savolainen et al., 2013)



Many genes of small effect or few genes of large effect?

The number of loci controlling a trait can affect the degree and frequency of local adaptation (Nuismer et al., 2007; Savolainen et al., 2013)

Percentage of phenotypic variance
in mycorrhizal traits attributed to
marker effects (R^2) ranged from
13% to 55%



Quantitative genetic analysis

- low genetic variation



SNP association analysis

- high genetic variation
-

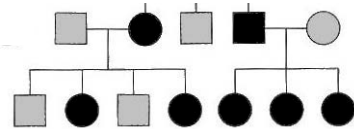
Quantitative genetic analysis

- low genetic variation



SNP association analysis

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Pedigree

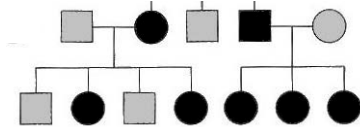


Relationship matrix

	A	B	C	D	E
A	1	.5	.25	0	.25
B		1	.5	.25	0
C			1	0	.25
D				1	.5
E					1

Quantitative genetic analysis

- low genetic variation



Pedigree

Relationship matrix

	A	B	C	D	E
A	1	.5	.25	0	.25
B		1	.5	.25	0
C			1	0	.25
D				1	.5
E					1

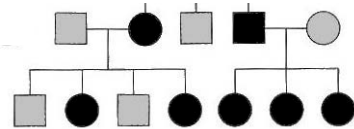
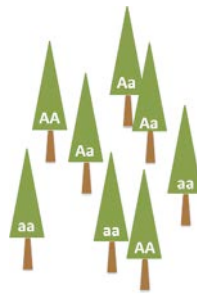


SNP association analysis

- high genetic variation

74517	A:G	A:G	G:G	A:A	A:A	G:G	C:C	A:G	G:G
74519	A:A	G:G	A:G	A:C	A:C	A:G	C:C	A:G	G:G
74521	A:A	G:G	A:G	A:C	A:C	A:G	A:C	A:G	A:G
74522	A:A	G:G	A:G	A:C	A:C	G:G	C:C	A:A	G:G
74612	A:G	A:G	G:G	A:A	A:C	A:G	C:C	A:A	G:G
74618	A:A	G:G	G:G	A:A	A:C	A:G	C:C	A:A	G:G
74623	A:G	A:G	G:G	A:A	A:A	G:G	C:C	A:A	G:G
74628	A:A	G:G	G:G	A:A	A:C	A:G	C:C	A:A	G:G
74711	A:A	G:G	G:G	A:A	A:C	G:G	C:C	A:G	G:G
74716	A:G	A:G	G:G	A:A	A:C	A:G	C:C	A:A	G:G

	A	B	C	D	E
A	1	.72	.31	.11	.21
B		1	.57	.42	.08
C			1	.10	.22
D				1	.47
E					1

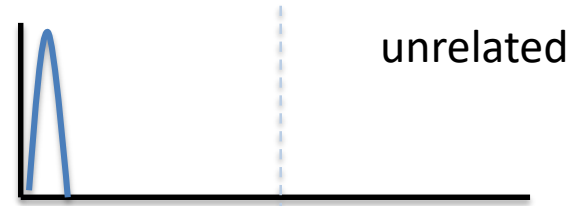


Pedigree

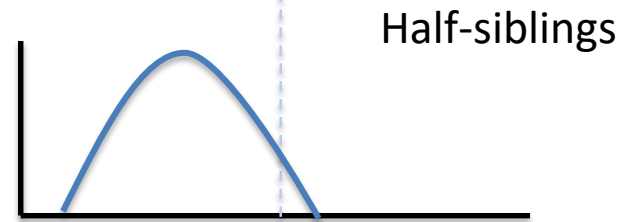


Relationship matrix

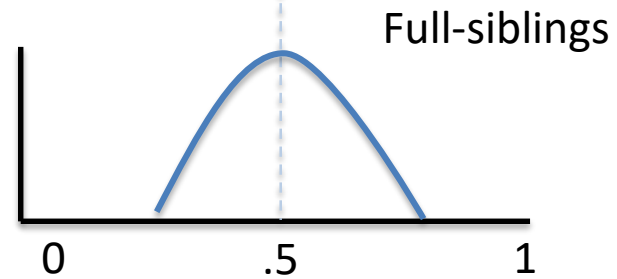
	A	B	C	D	E
A	1	.5	.25	0	.25
B		1	.5	.25	0
C			1	0	.25
D				1	.5
E					1



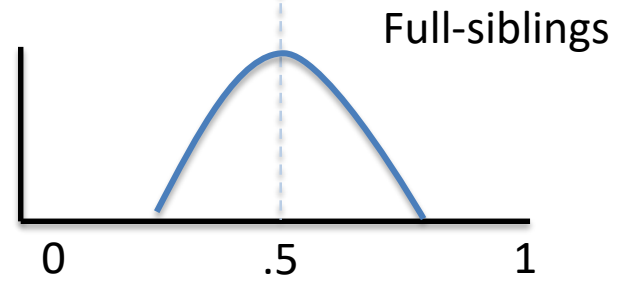
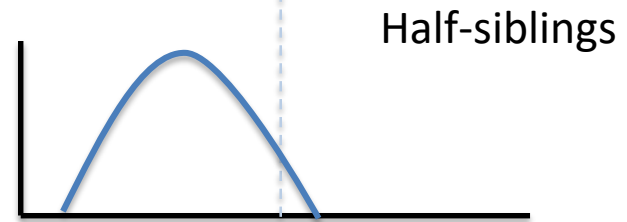
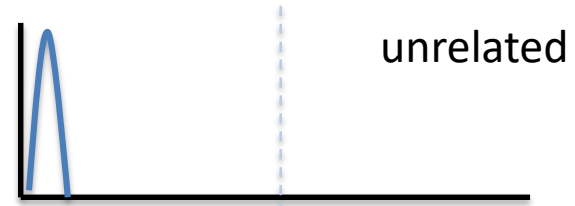
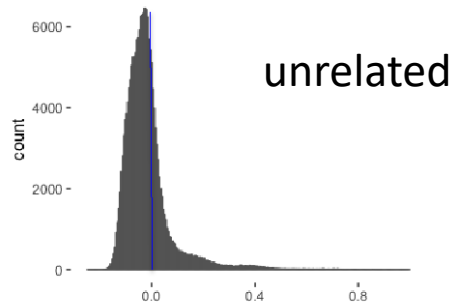
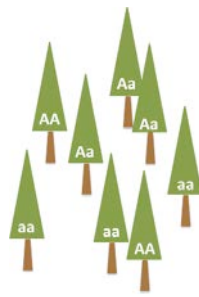
unrelated

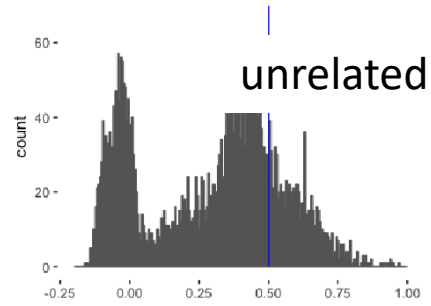
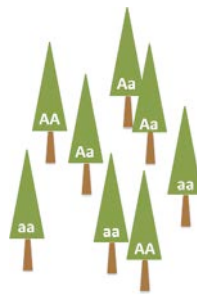


Half-siblings

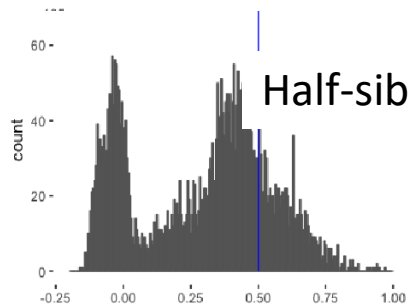


Full-siblings

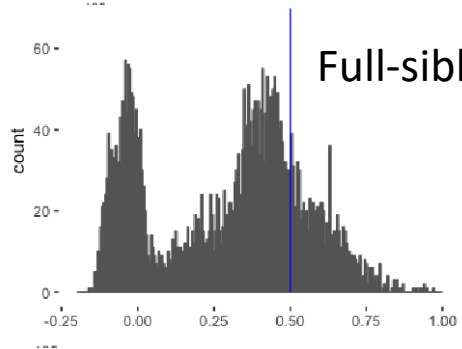




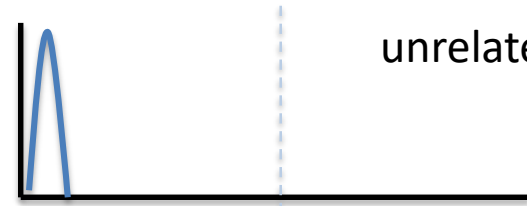
unrelated



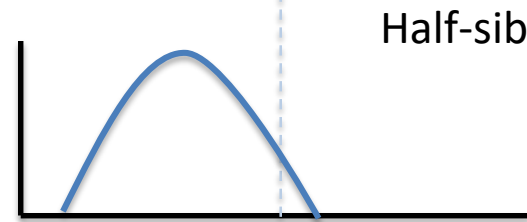
Half-siblings



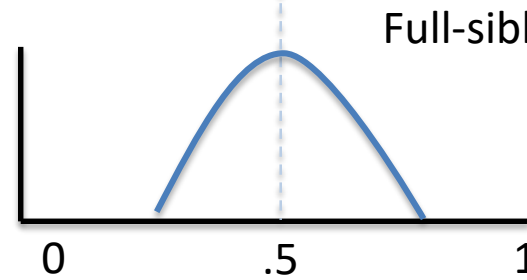
Full-siblings



unrelated



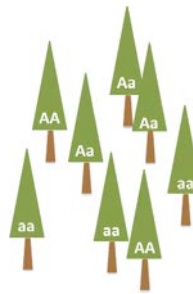
Half-siblings



Full-siblings

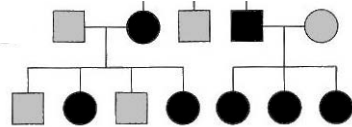
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74517	A:G	A:G	G:G	A:A	A:A	G:G	C:C	A:G	G:G
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74623	A:G	A:G	G:G	A:A	A:A	G:G	C:C	A:A	G:G
74628	A:A	G:G	G:G	A:A	A:C	A:G	C:C	A:A	G:G
74711	A:A	G:G	G:G	A:A	A:C	G:G	C:C	A:G	G:G
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B		1	.5	.25	0
C			1	0	.25
D				1	.5
E					1

	A	B	C	D	E
A	1	.72	.31	.11	.21
B		1	.57	.42	.08
C			1	.10	.22
D				1	.47
E					1

Chapter 3: Association mapping of ectomycorrhizal traits in loblolly pine

Q1: What can we say about the number and effect of genes involved in mycorrhizal traits in loblolly pine?

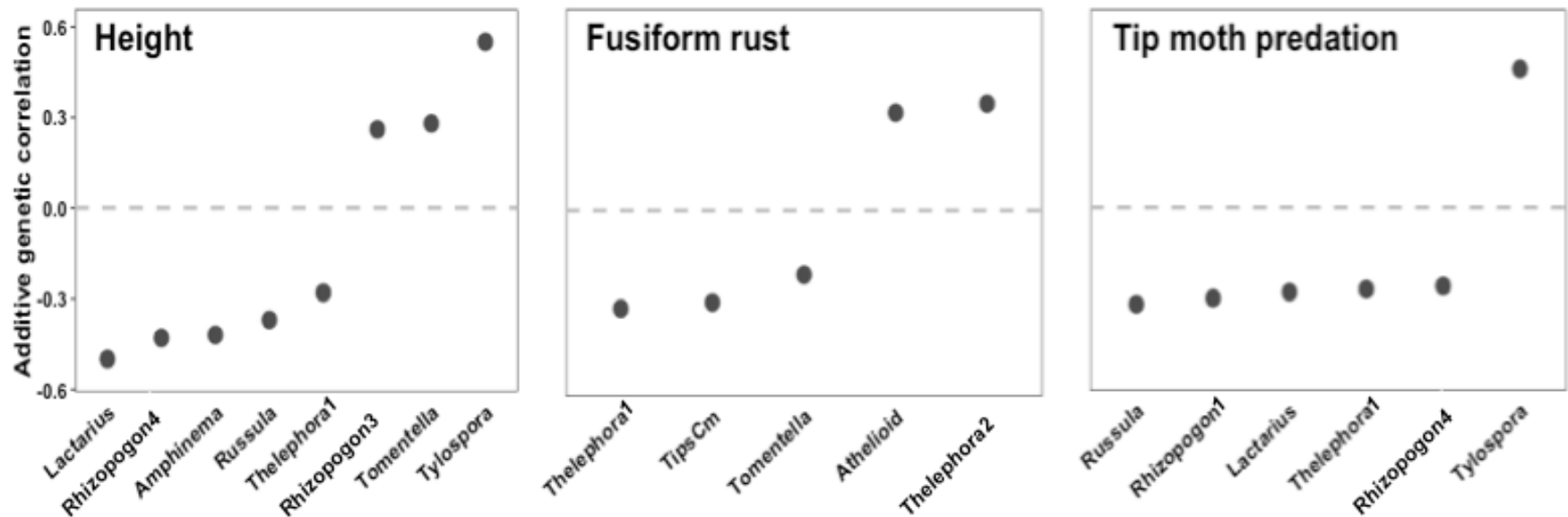
Q2: Can we determine a genetic basis for the correlations detected in the quantitative genetics study?

Q3: Can we ascribe putative function to any of the relevant SNPs?



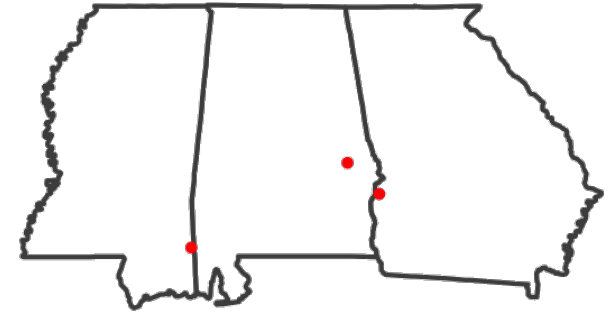
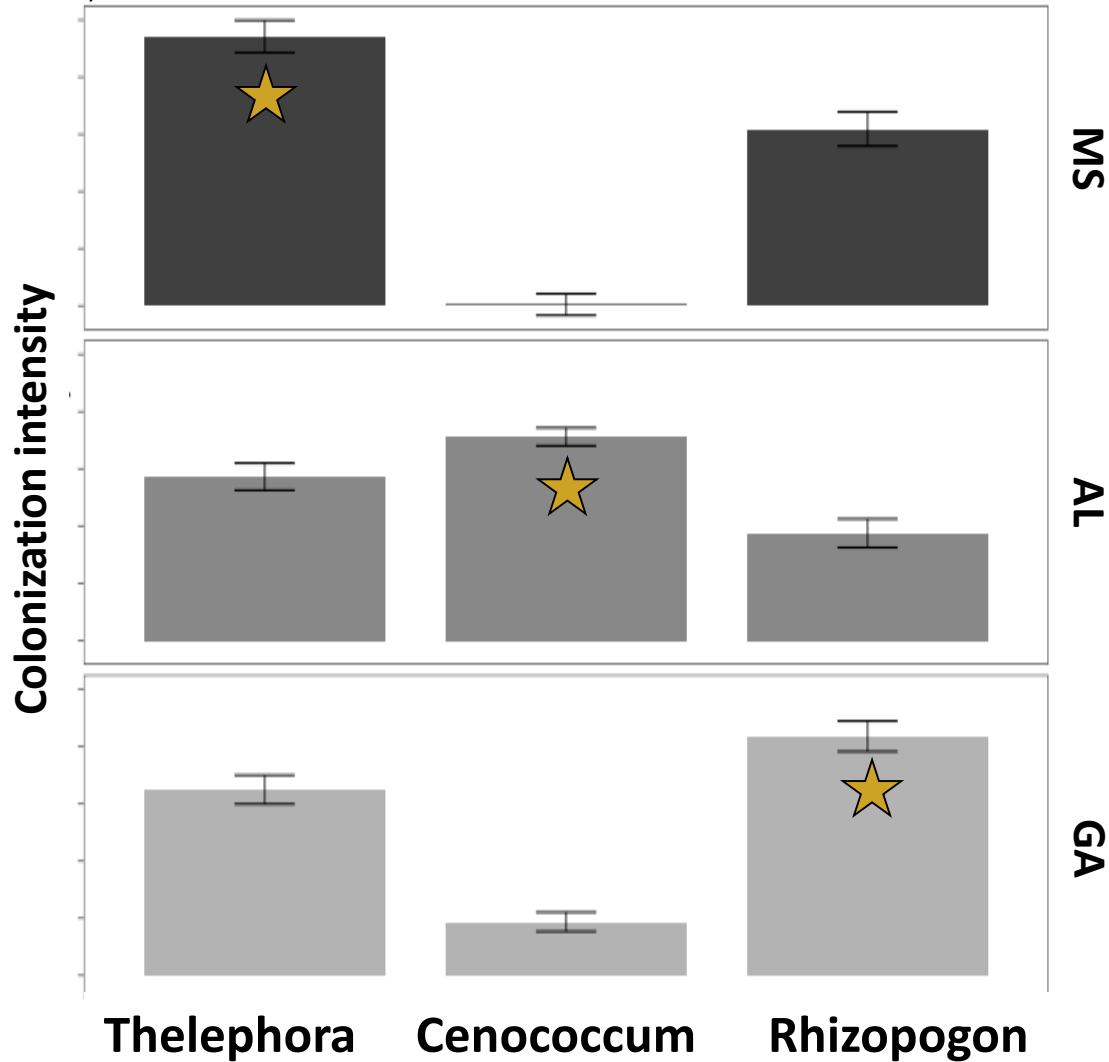
Can we use the SNP data to determine a probably genetic basis for the correlations detected in the quantitative genetics study?

Sadly, no.



The abundance of three of the four major fungal colonizers was determined solely by soil inoculation source

Rhizopogon ($F_{2,532} = 19.342$, $p = 7.787 \times 10^{-9}$), *Cenococcum* ($F_{2,532} = 120.840$, $p < 2.0 \times 10^{-16}$), and *Thelephora* ($F_{2,532} = 12.5084$, $p = 4.91 \times 10^{-6}$)



Comparison with Monterey Pine

(Hoeksema et al *Ecology* 2012)

- five sampling locations
- few dominant fungal sp
- differed among soil AND plant



Monterey Pine



Discussion



Monterey Pine

Both show

- variation in soil fungal community

Different population structures



Loblolly Pine

